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<p>(21) International Application Number: PCT/US97/19906</p> <p>(22) International Filing Date: 29 October 1997 (29.10.97)</p> <p>(30) Priority Data: 08/746,361 8 November 1996 (08.11.96) US</p> <p>(71) Applicant: IDEC PHARMACEUTICALS CORPORATION [US/US]; 11011 Torreyana Road, San Diego, CA 92121 (US).</p> <p>(72) Inventors: ANDERSON, Darrell, R.; 1851 Navajo Place, Escondido, CA 92029 (US). HANNA, Nabil; 3255 Fortuna Ranch Road, Olivenhain, CA 92024 (US). BRAMS, Peter; 4260 3rd Avenue #101, San Diego, CA 92103 (US).</p> <p>(74) Agents: GESS, E., Joseph et al.; Burns, Doane, Swecker & Mathis, L.L.P., P.O. Box 1404, Alexandria, VA 22313-1404 (US).</p>		<p>(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report.</i></p>	
<p>(54) Title: IDENTIFICATION OF UNIQUE BINDING INTERACTIONS BETWEEN CERTAIN ANTIBODIES AND THE HUMAN B7.1 AND B7.2 CO-STIMULATORY ANTIGENS</p> <p>(57) Abstract</p> <p>The present invention relates to the identification of antibodies which are specific to human B7.1 antigen (CD80) and which are capable of inhibiting the binding of B7.1 to a CD28 receptor and which are not capable of inhibiting the binding of B7.1 to a CTLA-4 receptor. Two of these antibodies, 16C10 and 7C10, significantly inhibit the production of IL-2, in spite of the existence of a second activating ligand B7.2 (CD86). Blocking of the primary activation signal between CD28 and B7.1 (CD80) with these antibodies while allowing the unimpaired or coincident interaction of CTLA-4 and B7.1 and/or B7.2 represents a combined antagonistic effect on positive co-stimulation with an agonistic effect on negative signalling. These antibodies may be used as specific immunosuppressants, e.g., for the treatment of autoimmune diseases and to prevent organ transplant rejection.</p>			

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IDENTIFICATION OF UNIQUE BINDING INTERACTIONS BETWEEN
CERTAIN ANTIBODIES AND THE HUMAN B7.1 AND B7.2
CO-STIMULATORY ANTIGENS

FIELD OF THE INVENTION

5 The present invention relates to the identification and use of monoclonal antibodies which are specific to B7.1 antigens (CD80). More specifically, the present invention relates to the identification and use of monoclonal antibodies or primatized forms thereof which are capable of inhibiting the binding of human B7.1 antigen to a CD28 receptor and which are not capable of inhibiting the binding of B7.1 to a CTLA-4 receptor. Thus, the invention relates to the identification and use of monoclonal antibodies and primatized forms thereof which recognize specific sites on the B7.1 antigen which are exclusive of CTLA-4 receptor binding.

10 The invention further relates to monoclonal antibodies or primatized forms thereof which recognize specific sites on the human B7.1 antigen and are capable of inhibiting IL-2 production.

15 Also, the present invention relates to pharmaceutical compositions containing monoclonal or primatized antibodies specific to human B7.1 and their use as immunosuppressants by modulating the B7:CD28 pathway, e.g., for the treatment of autoimmune disorders, and the prevention of organ rejection.

BACKGROUND OF THE INVENTION

The clinical interface between immunology, hematology, and oncology has long been appreciated.

20 Many conditions treated by the hematologist or oncologist have either an autoimmune or immunodeficient

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component to their pathophysiology that has led to the widespread adoption of immunosuppressive medications by hematologists, whereas oncologists have sought immunologic adjuvants that might enhance endogenous 5 immunity to tumors. To date, these interventions have generally consisted of nonspecific modes of immunosuppression and immune stimulation. In addition to the limited efficacy of these interventions, toxicities secondary to their nonspecificity have also 10 limited their overall success. Therefore, alternative strategies have been sought.

Elucidation of the functional role of a rapidly increasing number of cell surface molecules has contributed greatly to the integration of immunology 15 with clinical hematology and oncology. Nearly 200 cell surface antigens have been identified on cells of the immune and hematopoietic systems (Schlossman SF, Boumsell L, Gilks JM, Harlan T, Kishimoto, C Morimoto C, Ritz J., Shaw S, Silverstein RL, Springer TA, Tedder TF, 20 Todd RF:CD antigens (1993), Blood 83:879, 1994). These antigens represent both lineage-restricted and more widely distributed molecules involved in a variety of processes, including cellular recognition, adhesion, induction and maintenance of proliferation, cytokine 25 secretion, effector function, and even cell death. Recognition of the functional attributes of these molecules has fostered novel attempts to manipulate the immune response. Although molecules involved in cellular adhesion and antigen-specific recognition have 30 previously been evaluated as targets of therapeutic immunologic intervention, recent attention has focused on a subgroup of cell surface molecules termed co-stimulatory molecules (Bretscher P: "The two-signal model of lymphocyte activation twenty-one years later."

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Immunol. Today 13:73, (1992); Jenkins MK, Johnson JG: "Molecules involved in T-cell co-stimulation." Curr Opin Immunol 5:351, (1993); Geppert T, Davis L. Gur H. Wacholtz M. Lipsky P: "Accessory cell signals involved in T-cell activation." Immunol Rev 117:5, (1990); Weaver CT, Unanue ER: "The co-stimulatory function of antigen-presenting cells." Immunol Today 11:49, (1990); Stennam RM, Young JW: "Signals arising from antigen-presenting cells." Curr Opin Immunol 3:361, (1991)).

5 10 Co-stimulatory molecules do not initiate but rather enable the generation and amplification of antigen-specific T-cell responses and effector function (Bretscher P: "The two-signal model of lymphocyte activation twenty-one years later." Immunol. Today 13:73, (1992); Jenkins MK, Johnson JG: "Molecules involved in T-cell co-stimulation." Curr Opin Immunol 5:351, (1993); Geppert T, Davis L. Gur H. Wacholtz M. Lipsky P: "Accessory cell signals involved in T-cell activation." Immunol Rev 117:5, (1990); Weaver CT,

15 20 Unanue ER: "The co-stimulatory function of antigen-presenting cells." Immunol Today 11:49, (1990); Stennam RM, Young JW: "Signals arising from antigen-presenting cells." Curr Opin Immunol 3:361, (1991); June CH, Bluestone JA, Linsley PS, Thompson CD: "Role of the CD28 receptor in T-cell activation." Immunol Today 15:321, (1994)).

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Recently, one specific co-stimulatory pathway termed B7:CD28 has been studied by different research groups because of its significant role in B and T cell activation (June CH, Bluestone JA, Linsley PS, Thompson CD: "Role of the CD28 receptor in T-cell activation." Immunol Today 15:321, (1994); June CH, Ledbetter JA: "The role of the CD28 receptor during T-cell responses to antigen." Annu Rev Immunol 11:191, (1993); Schwartz

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RH: "Co-stimulation of T lymphocytes: The role of CD28, CTLA-4, and B7/BB1 in interleukin-2 production and immunotherapy." Cell 71:1065-1068, (1992); Jenkins MK, Taylor PS, Norton SD, Urdahl KB: "CD28 delivers a 5 costimulatory signal involved in antigen-specific IL-2 production by human T cells." Journal of Immunology 147:2461-2466 (1991). Since this ligand:receptor pathway was discovered four years ago, a large body of evidence has accumulated suggesting that B7:CD28 10 interactions represent one of the critical junctures in determining immune reactivity versus anergy (June CH, Bluestone JA, Linsley PS, Thompson CD: "Role of the CD28 receptor in T-cell activation." Immunol Today 15:321, (1994); June CH, Ledbetter JA: "The role of the CD28 receptor during T-cell responses to antigen." Annu Rev Immunol 11:191, (1993); Schwartz RH: "Co-stimulation of T lymphocytes: The role of CD28, CTLA-4, and B7/BB1 in interleukin-2 production and immunotherapy." Cell 71:1065-1068, (1992); Cohen J: "Mounting a targeted 15 strike on unwanted immune responses" (news; comment). Science 257:751, (1992); Cohen J: "New protein steals the show as 'co-stimulator' of T cells" (news; comment). Science 262:844, (1993)).

In particular, the role of the human B7 antigens, 20 i.e., human B7.1 (CD80) and B7.2 (CD86), has been reported to play a co-stimulatory role in T-cell activation. See, e.g., Gimmi CD, Freeman, GJ, Gribben JG, Sugita K, Freedman AS, Morimoto C, Nadler LM: "B-cell surface antigen B7 provides a costimulatory signal 25 that induces T cells to proliferate and secrete interleukin 2." Proc. Natl. Acad. Sci. (USA) 88:6575-6579 (1991).

1. B7.1 and B7.2 Co-stimulatory Role in T Cell Activation

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The elaboration of a successful immune response depends on a series of specific interactions between a T cell and an antigen presenting cell. Although the essential first step in this process depends upon the 5 binding of antigen to the T cell receptor, in the context of the MHC class II molecule (Lane, P.J.L., F.M. McConnell, G.L. Schieven, E.A. Clark, and J.A. Ledbetter, (1990), "The Role of Class II Molecules in Human B Cell Activation." The Journal of Immunology, 10 144:3684-3692), this interaction alone is not sufficient to induce all the events necessary for a sustained response to a given antigen (Schwartz, R.H. (1990), "A Cell Culture Model for T Lymphocyte Clonal Anergy." Science, 248:1349; Jenkins, M.K. (1992), "The Role of 15 Cell Division in the Induction of Clonal Anergy." Immunology Today, 13:69; Azuma, M., M. Cayabyab, D. Buck, J.H. Phillips, and L.L. Lanier, (1992), "Involvement of CD28 in MHC-unrestricted Cytotoxicity Mediated by a Human Natural Killer Leukemia Cell Line." 20 The Journal of Immunology, 149:1115-1123; Azuma, M., M. Cayabyab, D. Buck, J.H. Phillips, and L.L. Lanier, (1992), "CD28 Interaction with B7 Costimulates Primary Allogeneic Proliferative Responses and Cytotoxicity Mediated by Small Resting T Lymphocytes." J. Exp. Med., 25 175:353-360); S.D. Norton, L. Zuckerman, K.B. Urdahl, R. Shefner, J. Miller, and M.K. Jenkins, (1992), "The CD28 Ligand, B7, Enhances IL-2 Production by Providing a Costimulatory Signal to T Cells." The Journal of Immunology, 149:1556-1561; R. H. Schwartz, (1992), "Costimulation of T Lymphocytes: The Role of CD28, CTLA-4, and B7/BB1 in Interleukin-2 Production and 30 Immunotherapy." Cell 71:1065-1068).

The involvement of certain other co-stimulatory molecules is necessary (Norton, S.D., L. Zuckerman, K.B.

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Urdahl, R. Shefner, J. Miller, and M.K. Jenkins, (1992), "The CD28 Ligand, B7, Enhances IL-2 Production by Providing A Costimulatory Signal to T Cells." The Journal of Immunology, 149:1556-1561). "The homodimers 5 CD28 and CTLA-4 expressed on T cells" (June, C.H., J.A. Ledbetter, P.S. Linsley, and C.B. Thompson, (1990), "Role of the CD28 Receptor in T-Cell Activation." Immunology Today, 11:211-216; Linsley, P.S., W. Brady, M. Urnes, L.S. Grosmaire, N.K. Damle, and J.A. 10 Ledbetter, (1991), "CTLA-4 is a Second Receptor for the B Cell Activation Antigen B7." J. Exp. Med., 174:561), together with B7.1 (CD80) and B7.2 (CD86) expressed on antigen presenting cells, are major pairs of co-stimulatory molecules necessary for a sustained immune 15 response (Azuma, M., H. Yssel, J.H. Phillips, H. Spits, and L.L. Lanier, (1993), "Functional Expression of B7/BB1 on Activated T Lymphocytes." J. Exp. Med., 177:845-850; Freeman, G.J., A.S. Freedman, J.M. Segil, G. Lee, J.F. Whitman, and LM. Nadler, (1989), "B7, A 20 New Member of the Ig Superfamily with Unique Expression on Activated and Neoplastic B Cells." The Journal of Immunology, 143:2714-2722; Hathcock, K.S., G. Laslo, H.B. Dickler, J. Bradshaw, P. Linsley, and R.J. Hodes, (1993), "Identification of an Alternative CTLA-4 Ligand 25 Costimulatory for T Cell Activation." Science, 262:905-911; Hart, D.N.J., G.C. Starling, V.L. Calder, and N.S. Fernando, (1993). "B7/BB-1 is a Leucocyte Differentiation Antigen on Human Dendritic Cells Induced by Activation." Immunology, 79:616-620). It can be 30 shown in vitro that the absence of these co-stimulatory signals leads to an aborted T cell activation pathway and the development of unresponsiveness to the specific antigen, or anergy. (See, e.g., Harding, F.A., J.G. McArthur, J.A. Gross, D.M. Raulet, and J.P. Allison,

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(1992), "CD28 Mediated Signalling Co-simulates Murine T Cells and Prevents Induction of Anergy in T Cell Clones." Nature, 356:607-609; Gimmi, C.D., G.J. Freeman, J.G. Gribben, G. Gray, and L.M. Nadler, (1993); 5 "Human T-Cell Clonal Anergy is Induced by Antigen Presentation in the Absence of B7 Costimulation." Proc. Natl. Acad. Sci., 90:6586-6590; Tan, P., C. Anasefti, J.A. Hansen, J. Melrose, M. Brunvand, J. Bradshaw, J.A. Ledbetter, and P.S. Linsley, (1993), "Induction of 10 Alloantigen-specific Hyporesponsiveness in Human T Lymphocytes by Blocking Interaction of CD28 with Its Natural Ligand B7/BB1." J. Exp. Med., 177:165-173). Achievement of *in vivo* tolerance constitutes a mechanism for immunosuppression and a viable therapy for organ 15 transplant rejection and for the treatment of autoimmune diseases. This has been achieved in experimental models following the administration of CTLA-4Ig (Lenschow, D.J., Y. Zeng, R.J. Thistlethwaite, A. Montag, W. Brady, M.G. Gibson, P.S. Linsley, and J.A. Bluestone, (1992), 20 "Long-Term Survival of Xenogeneic Pancreatic Islet Grafts Induced by CTLA-4Ig." Science, 257:789-795). The molecules B7.1 and B7.2 can bind to either CD28 or CTLA-4, although B7.1 binds to CD28 with a Kd of 200 Nm and to CTLA-4 with a 20-fold higher affinity 25 (Linsley, P.S., E.A. Clark, and J.A. Ledbetter, (1990), "T-Cell Antigen CD28 Mediates Adhesion with B Cells by Interacting with Activation Antigen B7/BB-1." Proc. Natl. Acad. Sci., 87:5031-5035; Linsley et al, (1993), "The Role of the CD28 receptor during T cell responses 30 to antigen," Annu. Rev. Immunol., 11:191-192; Linesley et al, (1993), "CD28 Engagement by B7/BB-1 Induces Transient Down-Regulation of CD28 Synthesis and Prolonged Unresponsiveness to CD28 Signaling," The Journal of Immunology, 150:3151-3169). B7.1 is

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expressed on activated B cells and interferon induced monocytes, but not resting B cells (Freeman, G.J., G.S. Gray, C.D. Gimmi, D.B. Lomarrd, L-J. Zhou, M. White, J.D. Fingeroth, J.G. Gribben, and LM. Nadler, (1991). 5 "Structure, Expression and T Cell Costimulatory Activity of the Murine Homologue of the Human B Lymphocyte Activation Antigen B7," J. Exp. Med., 174:625-631). B7.2, on the other hand, is constitutively expressed at very low levels on resting monocytes, dendritic cells 10 and B cells, and its expression is enhanced on activated T cells, NK cells and B lymphocytes (Azuma, M. D. Ito, H. Yagita, K. Okumura, J.H. Phillips, L.L. Lanier, and C. Somoza, 1993, "B70 Antigen is a Second Ligand for CTLA-4 and CD28," Nature, 366:76-79). Although B7.1 and B7.2 can be expressed on the same cell type, their 15 expression on B cells occurs with different kinetics (Lenschow, D.J., G.H. Su, L.A. Zuckerman, N. Nabavi, C.L. Jellis, G.S. Gray, J. Miller, and J.A. Bluestone, (1993), "Expression and Functional Significance of an 20 Additional Ligand for CTLA-4," Proc. Natl. Acad. Sci., USA, 90:11054-11058; Boussiotis, V.A., G.J. Freeman, J.G. Gribben, J. Daley, G. Gray, and L.M. Nadler, (1993), "Activated Human B Lymphocytes Express Three 25 CTLA-4 Counter-receptors that Co-stimulate T-Cell Activation." Proc. Natl. Acad. Sci., USA, 90:11059-11063). Further analysis at the RNA level has demonstrated that B7.2 mRNA is constitutively expressed, whereas B7.1 mRNA is detected 4 hours after activation and initial low levels of B7.1 protein are not 30 detectable until 24 hours after stimulation (Boussiotis, V.A., G.J. Freeman, J.G. Gribben, J. Daley, G. Gray, and L.M. Nadler, (1993), "Activated Human B Lymphocytes Express Three CTLA-4 Counter-receptors that Co-stimulate T-Cell Activation," Proc. Natl. Acad. Sci., USA,

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90:11059-11063). CTLA-4/CD28 counter receptors, therefore, may be expressed at various times after B Cell activation.

More recently, it has been suggested that the 5 second T cell associated co-receptor CTLA-4 apparently functions as a negative modulator to override and prevent a runaway immune system (Krummel M, Allison J: "CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation." J. Exp. Med. 182:459-466 10 (1995)). The CTLA-4 receptor plays a critical role in down regulating the immune response, as evidenced in CTLA-4 knockout mice. Knockout mice born without the ability to express the CTLA-4 gene die within 3-4 weeks of severe lymphoproliferative disorder (Tivol EA, 15 Borriello G, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH: "Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4." Immunity 3:541-547 (1995)). CTLA-4 is 20 thought to function through signaling mechanisms linked to induction of apoptosis (Gribben JG, Freeman GJ, Boussiotis VA, Rennert P, Jellis CL, Greenfield E, Barber M, Restivo Jr. VA, Ke X, Gray GS, Nadler LM: "CTLA-4 mediates antigen specific apoptosis of human T 25 cells." Proc. Natl. Acad. Sci. (USA) 92:811-815 (1995)), triggered through as yet undefined ligand binding to specific cites on the receptor. It has been shown in vitro that the blocking of the B7.1/B7.2 dependent co-stimulatory signals in various ways leads to an aborted 30 T cell activating pathway and the development of unresponsiveness to the specific antigen (Lederman S, Chess L, Yellin MJ: "Murine monoclonal antibody (5c8) recognizes a human glycoprotein on the surface of T-lymphocytes, compositions containing same." U.S. Patent

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No. 5,474,771 (December 12, 1995); Linsley PS, Ledbetter JA, Damle NK, Brady W: "Chimeric CTLA4 receptor and methods for its use." U.S. Patent No. 5,434,131 (July 18, 1995); Harding, 1992; Gimmi CD, Freeman GJ, Bribben JG, Gray G, Nadler LM: "Human T-cell clonal anergy is induced by antigen presentation in the absence of B7 costimulation." Proc. Natl. Acad. Sci. (USA) 90:6586-6590 (1993); Tan P, Anasetti C, Hansen JA, Melrose J, Brunvand M, Bradshaw J, Ledbetter JA, Linsley PS: "Induction of alloantigen-specific hyporesponsiveness in human T lymphocytes by blocking interaction of CD28 with its natural ligand B7/BB1." J. Exp. Med. 177:165-173 (1993)). Achievement of in vivo tolerance, anergy, or depleting of antigen-specific T cells would constitute a mechanism for immunosuppression and a viable therapy for organ transplant rejection or plausible treatment for autoimmune diseases.

The differential temporal expression of B7.1 and B7.2 suggests that the interaction of these two molecules with CTLA-4 and/or CD28 deliver distinct but related signals to the T cell (LaSalle, J.M., P.J. Tolentino, G.J. Freeman, L.M. Nadler, and D.A. Hafler, (1992), "CD28 and T Cell Antigen Receptor Signal Transduction Coordinately Regulate Interleukin 2 Gene Expression In Response to Superantigen Stimulation," J. Exp. Med., 176:177-186; Vandenberghe, P., G.J. Freeman, L.M. Nadler, M.C. Fletcher, M. Kamoun, L.A. Turka, J.A. Ledbetter, C.B. Thompson, and C.H. June, (1992), "Antibody and B7/BB1-mediated Ligation of the CD28 Receptor Induces Tyrosine Phosphorylation in Human T Cells," The Journal of Experimental Medicine, 175:951-960). The exact signaling functions of CTLA-4 and CD28 on the T cell are currently unknown (Janeway, C.A., Jr. and K. Bottomly, (1994), "Signals and Signs

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for Lymphocyte Responses," Cell, 76:275-285). However, it is possible that one set of receptors could provide the initial stimulus for T cell activation and the second, a sustained signal to allow further elaboration 5 of the pathway and clonal expansion to take place (Linsley, P.S., J.L. Greene, P. Tan, J. Bradshaw, J.A. Ledbetter, C. Anasetti, and N.K. Damle, (1992), "Coexpression and Functional Cooperation of CTLA-4 and CD28 on Activated T Lymphocytes," J. Exp. Med., 10 176:1595-1604). The current data supports the two-signal hypothesis proposed by Jenkins and Schwartz (Schwartz, R.H., (1990), "A Cell Culture Model for T Lymphocyte Clonal Anergy," Science, 248:1349; Jenkins, M.K., (1992), "The Role of Cell Division in the 15 Induction of Clonal Anergy," Immunology Today, 13:69) that both a TCR and co-stimulatory signal are necessary for T cell expansion, lymphokine secretion and the full development of effector function (Greenan, V. and G. Kroemer, (1993), "Multiple Ways to Cellular Immune 20 Tolerance," Immunology Today, 14:573). The failure to deliver the second signal results in the inability of T cells to secrete IL-2 and renders the cell unresponsive to antigen.

Structurally, both B7.1 and B7.2 contain 25 extracellular immunoglobulin superfamily V and C-like domains, a hydrophobic transmembrane region and a cytoplasmic tail (Freeman, G.J., J.G. Gribben, V.A. Boussiotis, J.W. Ng, V. Restivo, Jr., L.A. Lombard, G.S. Gray, and L.M. Nadler, (1993), "Cloning of B7.2: 30 A CTLA-4 Counter-receptor that Co-stimulates Human T Cell Proliferation," Science, 262:909). Both B7.1 and B7.2 are heavily glycosylated. B7.1 is a 44-54kD glycoprotein comprised of a 223 amino acid extracellular domain, a 23 amino acid transmembrane domain, and a 61

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amino acid cytoplasmic tail. B7.1 contains 3 potential protein kinase phosphorylation sites. (Azuma, M., H. Yssel, J.H. Phillips, H. Spits, and L.L. Lanier, (1993), "Functional Expression of B7/BB1 on Activated T Lymphocytes," J. Exp. Med., 177:845-850). B7.2 is a 306 amino acid membrane glycoprotein. It consists of a 220 amino acid extracellular region, a 23 amino acid hydrophobic transmembrane domain and a 60 amino acid cytoplasmic tail (Freeman, G.J., A.S. Freedman, J.M. Segil, G. Lee, J.F. Whitman, and LM. Nadler, (1989), "B7, A New Member of the Ig Superfamily with Unique Expression on Activated and Neoplastic B Cells," The Journal of Immunology, 143:2714-2722). Although both B7.1 and B7.2 genes are localized in the same chromosomal region (Freeman, G.J., D.B. Lombard, C.D. Gimmi, S.A. Brod, L Lee, J.C. Laning, D.A. Hafler, M.E. Dorf, G.S. Gray, H. Reiser, C.H. June, C.B. Thompson, and L.M. Nadler, (1992), "CTLA-4 and CD28 mRNA are Coexpressed in Most T Cells After Activation," The Journal of Immunology, 149:3795-3801; Schwartz, R.H., (1992), "Costimulation of T Lymphocytes: The Role of CD28, CTLA-4, and B7/BB1" in Selvakumar, A., B.K. Mohanraj, R.L. Eddy, T.B. Shows, P.C. White, C. Perrin, and B. Dupont, (1992), "Genomic Organization and Chromosomal Location of the Human Gene Encoding the B-Lymphocyte Activation Antigen B7," Immunogenetics, 36:175-181), these antigens do not share a high level of homology. The overall homology between B7.1 and B7.2 is 26% and between murine B7.1 and human B7.1 is 27% (Azuma, M., H. Yssel, J.H. Phillips, H. Spits, and L.L. Lanier, (1993), "Functional Expression of B7/BB1 on Activated T Lymphocytes," J. Exp. Med., 177:845-850; Freeman, G.J., A.S. Freedman, J.M. Segil, G. Lee, J.F. Whitman, and LM. Nadler, (1989), "B7, A New Member of

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the Ig Superfamily with Unique Expression on Activated and Neoplastic B Cells," The Journal of Immunology, 143:2714-2722). Although alignment of human B7.1 human B7.2 and murine B7.1 sequences shows few stretches of 5 lengthy homology, it is known that all three molecules bind to human CTLA-4 and CD28. Thus, there is most likely a common, or closely homologous region shared by the three molecules that may be either contiguous or conformational. This region may constitute the binding 10 site of the B7.1 and B7.2 molecules to their counter-receptors. Antibodies raised against these epitopes could potentially inhibit the interaction of B7 with its counter-receptor on the T cell. Furthermore, antibodies that cross-reacted with this region on both B7.1 and 15 B7.2 molecules would potentially have practical advantages over antibodies directed against B7.1 or B7.2 separately.

2. Blockade of the B7/CD28 Interaction

Blocking of the B7/CD28 interaction offers the 20 possibility of inducing specific immunosuppression, with potential for generating long lasting antigen-specific therapeutic effects. Antibodies or agents that temporarily prevent this interaction may be useful, specific and safe clinical immunosuppressive agents, 25 with potential for generating long term antigen-specific therapeutic effects.

Antibodies to either B7.1 or B7.2 have been shown to block T cell activation, as measured by the inhibition of IL-2 production in vitro (DeBoer, M., P. 30 Parren, J. Dove, F. Ossendorp, G. van der Horst, and J. Reeder, (1992), "Functional Characterization of a Novel Anti-B7 Monoclonal Antibody," Eur. Journal of Immunology, 22:3071-3075; Azuma, M., H. Yssel, J.H.

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Phillips, H. Spits, and L.L. Lanier, (1993), "Functional Expression of B7/BB1 on Activated T Lymphocytes," J. Exp. Med., 177:845-850). However, different antibodies have been shown to vary in their immunosuppressive 5 potency, which may reflect either their affinity or epitope specificity. A possible explanation for this may reside in the ability of some antibodies to block only the binding of B7 to CD28, while promoting apoptosis or some other form of negative signaling 10 through the CTLA-4 receptor in activated T cells. Some antibodies to B7.1 or B7.2 may, in fact, hinder the activity of CTLA-4 by cross-reacting with the CTLA-4 binding domain. CTLA-4Ig fusion protein and anti-CD28 Fabs were shown to have similar effects on the down 15 regulation of IL-2 production.

In vivo administration of a soluble CTLA-4Ig fusion protein has been shown to suppress T cell dependent antibody responses in mice (Linsley, P.S., J.L. Greene, P. Tan, J. Bradshaw, J.A. Ledbetter, C. Anasetti, and 20 N.K. Damle, (1992), "Coexpression and Functional Cooperation of CTLA-4 and CD28 on Activated T Lymphocytes," J. Exp. Med., 176:1595-1604; Lin, H., S.F. Builing, P.S. Linsley, R.O. Wei, C.D. Thompson, and L.A. Turka, (1993), "Long-term Acceptance of Major 25 Histocompatibility Complex Mismatched Cardiac Allografts Induced by CTLA-4-Ig Plus Donor Specific Transfusion," J. Exp. Med., 178:1801) and, furthermore, larger doses were also able to suppress responses to a second immunization, demonstrating the feasibility of this 30 approach for the treatment of antibody mediated autoimmune disease. In addition, CTLA-4Ig was able to prevent pancreatic islet cell rejection in mice by directly inhibiting the interaction of T cells and B7.1/B7.2 antigen presenting cells (Lenschow, D.J., G.H.

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Su, L.A. Zuckerman, N. Nabavi, C.L. Jellis, G.S. Gray, J. Miller, and J.A. Bluestone, (1993), "Expression and Functional Significance of an Additional Ligand for CTLA-4," Proc. Natl. Acad. Sci., USA, 90:11054-11058).

5 In this case, long term donor specific tolerance was achieved.

3. Recombinant Phage Display Technology for Antibody Selection

To date, no monoclonal antibodies which crossreact
10 with both B7.1 and B7.2 have been reported. Furthermore, no monoclonal antibodies which are specific to B7.1 or B7.2 and which also recognize specific sites on the antigens which are restricted to co-activation receptor CD28 binding have been reported. Or
15 alternatively, no monoclonal antibodies which are specific to B7.1 or B7.2 and which recognize specific sites on the antigens which are exclusive of CTLA-4 receptor binding have been reported. As discussed supra, such antibodies would potentially be highly
20 desirable as immunosuppressants.

Phage display technology is beginning to replace traditional methods for isolating antibodies generated during the immune response, because a much greater percentage of the immune repertoire can be assessed than
25 is possible using traditional methods. This is in part due to PEG fusion inefficiency, chromosomal instability, and the large amount of tissue culture and screening associated with heterohybridoma production. Phage display technology, by contrast, relies on molecular techniques for potentially capturing the entire
30 repertoire of immunoglobulin genes associated with the response to a given antigen.

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This technique is described by Barbas et al, Proc. Natl. Acad. Sci., USA, 88, 7978-7982, (1991). Essentially, immunoglobulin heavy chain genes are PCR amplified and cloned into a vector containing the gene 5 encoding the minor coat protein of the filamentous phage M13 in such a way that a heavy chain fusion protein is created. The heavy chain fusion protein is incorporated into the M13 phage particle together with the light chain genes as it assembles. Each recombinant phage 10 contains, within its genome, the genes for a different antibody Fab molecule which it displays on its surface. Within these libraries, in excess of 10^6 different antibodies can be cloned and displayed. The phage library is panned on antigen coated microliter wells, 15 non-specific phage are washed off, and antigen binding phage are eluted. The genome from the antigen-specific clones is isolated and the gene III is excised, so that antibody can be expressed in soluble Fab form for further characterization. Once a single Fab is selected 20 as a potential therapeutic candidate, it may easily be converted to a whole antibody. A previously described expression system for converting Fab sequences to whole antibodies is IDEC's mammalian expression vector NEOSPLA. This vector contains either human gamma 1 or 25 gamma 4 constant region genes. CHO cells are transfected with the NEOSPLA vectors and after amplification this vector system has been reported to provide very high expression levels (> 30 pg/cell/day) can be achieved.

30 4. Primatized Antibodies

Another highly efficient means for generating recombinant antibodies is disclosed by Newman, (1992), Biotechnology, 10, 1455-1460. More particularly, this

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technique results in the generation of primatized antibodies which contain monkey variable domains and human constant sequences. This reference is incorporated by reference in its entirety herein.

5 Moreover, this technique is also described in commonly assigned U.S. Application No. 08/379,072, filed on January 25, 1995, which is a continuation of U.S. Serial No. 07/912,292, filed July 10, 1992, which is a continuation-in-part of U.S. Serial No. 07/856,281, 10 filed March 23, 1992, which is finally a continuation-in-part of U.S. Serial No. 07/735,064, filed July 25, 1991. 08/379,072 and the parent application thereof are incorporated by reference in their entirety herein.

This technique modifies antibodies such that they 15 are not antigenically rejected upon administration in humans. This technique relies on immunization of cynomolgus monkeys with human antigens or receptors. This technique was developed to create high affinity monoclonal antibodies directed to human cell surface 20 antigens.

Identification of macaque antibodies to human B7.1 and B7.2 by screening of phage display libraries or monkey heterohybridomas obtained using B lymphocytes from B7.1 and/or B7.2 immunized monkeys is also 25 described in commonly assigned U.S. Application No. 08/487,550, filed June 7, 1995, incorporated by reference in its entirety herein. More specifically, 08/487,550 provides four monoclonal antibodies 7B6, 16C10, 7C10 and 20C9 which inhibit the B7:CD28 pathway 30 and thereby function as effective immunosuppressants.

Antibodies generated in the manner described by these co-assigned applications have previously been reported to display human effector function, have reduced immunogenicity, and long serum half-life. The

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technology relies on the fact that despite the fact that cynomolgus monkeys are phylogenetically similar to humans, they still recognize many human proteins as foreign and therefore mount an immune response.

5 Moreover, because the cynomolgus monkeys are phylogenetically close to humans, the antibodies generated in these monkeys have been discovered to have a high degree of amino acid homology to those produced in humans. Indeed, after sequencing macaque 10 immunoglobulin light and heavy chain variable region genes, it was found that the sequence of each gene family was 85-98% homologous to its human counterpart (Newman et al, (1992), Id.). The first antibody generated in this way, an anti-CD4 antibody, was 91-92% 15 homologous to the consensus sequence of human immunoglobulin framework regions. Newman et al, Biotechnology, 10:1458-1460, (1992).

Monoclonal antibodies specific to the human B7 antigen have been previously described in the 20 literature. For example, Weyl et al, Hum. Immunol., 31(4), 271-276, (1991) describe epitope mapping of human monoclonal antibodies against HLA-B-27 using natural and mutated antigenic variants. Also, Toubert et al, Clin. Exp. Immunol., 82(1), 16-20, (1990) describe epitope 25 mapping of an HLA-B27 monoclonal antibody that also reacts with a 35-KD bacterial outer membrane protein. Also, Valle et al, Immunol., 69(4), 531-535, (1990) describe a monoclonal antibody of the IgG1 subclass which recognizes the B7 antigen expressed in activated B 30 cells and HTLV-1-transformed T cells. Further, Toubert et al, J. Immunol., 141(7), 2503-9, (1988) describe epitope mapping of HLA-B27 and HLA-B7 antigens using intradomain recombinants constructed by making hybrid genes between these two alleles in *E. coli*.

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High expression of B7 antigen has been correlated to autoimmune diseases by some researchers. For example, Ionesco-Tirgoviste et al, Med. Interne, 24(1), 11-17, (1986) report increased B7 antigen expression in 5 type 1 insulin-dependent diabetes. Also, the involvement of B7 antigen expression on dermal dendritic cells obtained from psoriasis patients has been reported. (Nestle et al, J. Clin. Invest., 94(1), 202-209, (1994)).

10 Further, the inhibition of anti-HLA-B7 alloreactive CTL using affinity-purified soluble HLA-B7 has been reported in the literature. (Zavazava et al, Transplantation, 51(4), 838-42, (1991)). Further, the use of B7 receptor soluble ligand, CTLA-4-Ig to block B7 15 activity (See, e.g., Lenschow et al, Science, 257, 789, 7955 (1992)) in animal models and a B7.1-Ig fusion protein capable of inhibiting B7 has been reported.

20 Evidence is provided in this disclosure for the identification of monoclonal antibodies which recognize specific sites on the B7.1 antigen which are restricted to CD28 receptor binding. Furthermore, evidence is presented herein for the identification of antibodies which recognize sites on the B7.1 antigen which are exclusive of CTLA-4 receptor binding. Thus, evidence is 25 presented herein to support the existence of unique antigen binding sites on the human B7.1 (CD80) co-stimulatory antigen. The sites claimed are identified by anti-B7.1 PRIMATIZED® antibodies and evidence is presented which confirms binding to a site of 30 interaction on the B7.1 antigen which is restricted to binding with the co-activation receptor CD28.

SUMMARY AND OBJECTS OF THE INVENTION

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An object of the invention is to identify novel antibodies which are specific to human B7.1 antigen. More specifically, it is an object of the invention to identify antibodies which are specific to human B7.1 antigen and which are also capable of inhibiting the binding of B7.1 to a CD28 receptor. It is also an object of this invention to identify antibodies which are specific to human B7.1 antigen and which are not capable of inhibiting the binding of B7.1 to a CTLA-4 receptor. Thus, an object of this invention is to identify antigens which recognize specific sites on the B7.1 antigen, wherein the recognized sites are restricted to CD28 receptor binding and which are exclusive of CTLA-4 receptor binding.

It is a further object of the invention to identify antibodies which are specific to human B7.1 antigen and which fail to recognize human B7.2 antigen.

It is another object of the invention to identify monoclonal antibodies and primatized forms thereof which recognize specific sites on the human B7.1 antigen and which inhibit IL-2 production and T cell proliferation and which function as effective immunosuppressants. More specifically, it is an object of this invention to identify antibodies which are specific to B7.1 and which are capable of inhibiting IL-2 production.

It is another object of the invention to provide monoclonal antibodies and primatized forms thereof which inhibit antigen driven responses in donor spleen cell cultures, e.g., antigen specific IgG responses, IL-2 production and cell proliferation.

It is another specific object of the invention to identify particular monoclonal antibodies specific to human B7.1 antigen and primatized forms thereof having advantageous properties, i.e., affinity,

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immunosuppressive activity, which are useful as therapeutics. More specifically, these antibodies and primatized forms thereof are to be used, e.g., as immunosuppressants, i.e., to block antigen driven immune responses, to treat autoimmune diseases such as 5 psoriasis, rheumatoid arthritis, systemic erythematosus (SLE), type 1 diabetes mellitus, idiopathic thrombocytopenia purpura (ITP), allergy, inflammatory bile disease, and to prevent organ rejection.

10 It is another object of the invention to provide pharmaceutical compositions containing one or more monoclonal antibodies specific to human B7.1 antigen or primatized forms thereof, and a pharmaceutically acceptable carrier or excipient. These compositions 15 will be used, e.g., as immunosuppressants to treat autoimmune diseases, e.g., idiopathic thrombocytopenia purpura (ITP) and systemic lupus erythematosus (SLE), to block antigen driven immune responses, and to prevent organ rejection in transplant recipients.

20 It is another object of the invention to provide novel methods of therapy by administration of therapeutically effective amounts of one or more or primatized monoclonal antibodies which specifically bind to human B7.1 antigen. Such therapeutic methods are 25 useful for treatment of diseases treatable by inhibition of the B7:CD28 pathway, e.g., autoimmune diseases such as idiopathic thrombocytopenia purpura (ITP), systemic lupus erythematosus (SLE), type 1 diabetes mellitus, psoriasis, rheumatoid arthritis, multiple sclerosis, 30 aplastic anemia, as well as for preventing rejection in transplantation subjects.

It is still another object of the invention to provide transfectants, e.g., CHO cells, which express at

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least the variable heavy and light domains of monoclonal antibodies specific to the human B7.1 antigen.

Definitions

The following terms are defined so that the invention may be more clearly understood.

5 Depleting antibody - an antibody which kills activated B cells or other antigen presenting cells.

10 Non-depleting antibody - an antibody which blocks the co-stimulatory action of B7 and T cell activating ligands CD28 and CTLA-4. Thus, it anergizes but does not eliminate the antigen presenting cell.

15 Primatized antibody - a recombinant antibody which has been engineered to contain the variable heavy and light domains of a monkey antibody, in particular, a cynomolgus monkey antibody, and which contains human constant domain sequences, preferably the human immunoglobulin gamma 1 or gamma 4 constant domain (or PE variant). The preparation of such antibodies is described in Newman et al, (1992), "Primatization of

20 Recombinant Antibodies for Immunotherapy of Human Diseases: A Macaque/Human Chimeric Antibody Against Human CDH, Biotechnology, 10:1458-1460; also in commonly assigned 08/379,072 both of which are incorporated by reference in their entirety herein. These antibodies

25 have been reported to exhibit a high degree of homology to human antibodies, i.e., 85-98%, display human effector functions, have reduced immunogenicity, and may exhibit high affinity to human antigens.

30 B7 antigens - B7 antigens in this application include, e.g., human B7, B7.1 and B7.2 antigens. These antigens bind to CD28 and/or CTLA-4. These antigens have a co-stimulatory role in T cell activation. Also, these B7 antigens all contain extracellular immunoglobulin

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superfamily V and C-like domains, a hydrophobic transmembrane region and a cytoplasmic tail. (See, Freeman et al, Science, 262:909, (1993)), and are heavily glycosylated.

5 Anti-B7 antibodies - Antibodies, preferably monkey monoclonal antibodies or primatized forms thereof, which specifically bind human B7 antigens, e.g., human B7.1 and/or B7.2 antigen with a sufficient affinity to block the B7:CD28 interaction and thereby induce
10 immunosuppression.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 depicts the pMS vector used to screen recombinant immunoglobulin libraries produced against B7 displayed on the surface of filamentous phage which
15 contains primers based on macaque immunoglobulin sequences.

Figure 2 depicts the NEOSPLA expression vector used to express the subject primatized antibodies specific to human B7.1 antigen.

20 Figure 3a depicts the amino acid and nucleic acid sequence of a primatized form of the light chain of 7C10.

Figure 3b depicts the amino acid and nucleic acid sequence of a primatized form of the heavy chain of
25 7C10.

Figure 4a depicts the amino acid and nucleic acid sequence of a primatized form of the light chain of 7B6.

Figure 4b depicts the amino acid and nucleic acid sequence of a primatized form of the heavy chain of 7B6.

30 Figure 5a depicts the amino acid and nucleic acid sequence of a primatized light chain 16C10.

Figure 5b depicts the amino acid and nucleic acid sequence of a primatized heavy chain 16C10.

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Figure 6 depicts the inability of P16C10 to block CTLA-4Ig-Biotin binding to B7.1 transfected CHO cells.

Figure 7 depicts the inability of CTLA-4Ig to block P16C10-Biotin binding to B7.1 transfected CHO cells.

5 Figure 8 depicts that BB-1 completely blocks binding of CTLA-4Ig-Biotin to B7.1 transfected CHO cells and further depicts the inability of BB-1 to significantly affect P16C10-Biotin binding to B7.1 transfected CHO cells.

10 Figure 9 depicts that CTLA-4Ig-Biotin is effectively blocked by all B7.1 inhibitors except P16C10.

15 Figure 10 depicts the ability of P16C10 to block binding of the CD28/B7-1Ig interaction. Data shown are averages of values obtained from four separate experiments.

DETAILED DESCRIPTION OF THE INVENTION

As described above, the present invention relates to the identification of monoclonal antibodies or 20 primatized forms thereof which are specific to human B7.1 antigen and which are capable of inhibiting the binding of B7.1 to a CD28 receptor and which are not capable of inhibiting the binding of B7.1 to a CTLA-4 receptor. Blocking of the primary activation site 25 between CD28 and B7.1 (CD80) with the identified antibodies while allowing the combined antagonistic effect on positive co-stimulation with an agnostic effect on negative signaling will be a useful therapeutic approach for intervening in relapsed forms 30 of autoimmune disease. The functional activity of the identified antibodies is defined by blocking the production of the T cell stimulatory cytokine IL-2. Identified antibodies have demonstrated the ability to

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block the production of IL-2 in excess of 50%, in spite of the existence of a second actuating ligand B7.2, suggesting an alternate mechanism of action exists which is not typical of the observed effects of other anti-
5 B7.1 antibodies defined in the literature.

Manufacture of novel monkey monoclonal antibodies which specifically bind human B7.1 and/or human B7.2 antigen, as well as primatized antibodies derived therefrom is described in co-pending U.S. Application 10 Serial No. 08/487,550, and as set forth herein. These antibodies possess high affinity to human B7.1 and/or B7.2 and therefore may be used as immunosuppressants which inhibit the B7:CD86 pathway.

Preparation of monkey monoclonal antibodies will 15 preferably be effected by screening of phage display libraries or by preparation of monkey heterohybridomas using B lymphocytes obtained from B7 (e.g., human B7.1 and/or B7.2) immunized monkeys.

As noted, the first method for generating anti-B7 20 antibodies involves recombinant phage display technology. This technique is generally described *supra*.

Essentially, this will comprise synthesis of 25 recombinant immunoglobulin libraries against B7 antigen displayed on the surface of filamentous phage and selection of phage which secrete antibodies having high affinity to B7.1 and/or B7.2 antigen. As noted *supra*, preferably antibodies will be selected which bind to both human B7.1 and B7.2. To effect such methodology, 30 the present inventors have created a unique library for monkey libraries which reduces the possibility of recombination and improves stability. This vector, PMS, is described in detail *infra*, and is shown in Figure 1.

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Essentially, to adopt phage display for use with macaque libraries, this vector contains specific primers for PCR amplifying monkey immunoglobulin genes. These primers are based on macaque sequences obtained while 5 developing the primatized technology and databases containing human sequences.

Suitable primers are disclosed in commonly assigned 08/379,072 incorporated by reference herein.

The second method involves the immunization of 10 monkeys, i.e., macaques, against human B7 antigen, preferably against human B7.1 and B7.2 antigen. The inherent advantage of macaques for generation of monoclonal antibodies is discussed *supra*. In particular, such monkeys, i.e., cynomolgus monkeys, may 15 be immunized against human antigens or receptors. Moreover, the resultant antibodies may be used to make primatized antibodies according to the methodology of Newman et al, Biotechnology, 10, 1455-1460, (1992), and Newman et al, commonly assigned U.S. Serial No. 20 08/379,072, filed January 25, 1995, which are incorporated by reference in their entirety.

The significant advantage of antibodies obtained 25 from cynomolgus monkeys is that these monkeys recognize many human proteins as foreign and thereby provide for the formation of antibodies, some with high affinity to desired human antigens, e.g., human surface proteins and cell receptors. Moreover, because they are phylogenetically close to humans, the resultant antibodies exhibit a high degree of amino acid homology to those 30 produced in humans. As noted above, after sequencing macaque immunoglobulin light and heavy variable region genes, it was found that the sequence of each gene family was 85-88% homologous to its human counterpart (Newman et al, (1992), Id.).

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Essentially, cynomolgus macaque monkeys are administered human B7 antigen, e.g., human B7.1 and/or human B7.2 antigen, B cells are isolated therefrom, e.g., lymph node biopsies are taken from the animals, 5 and B lymphocytes are then fused with KH6/B5 (mouse x human) heteromyeloma cells using polyethylene glycol (PEG). Heterohybridomas secreting antibodies which bind human B7 antigen, e.g., human B7.1 and/or human B7.2 antigen, are then identified.

10 Antibodies which bind to both B7.1 and B7.2 are desirable because such antibodies potentially may be used to inhibit the interaction of B7.1 and B7.2, as well as B7 with their counter-receptors, i.e., human CTLA-4 and CD28. Antibodies against these epitopes may 15 inhibit the interaction of both human B7.1 and human B7.2 with their counter receptors on the T cell. This may potentially provide synergistic effects.

20 However, antibodies which bind to only one of human B7 antigen, B7.1 antigen or B7.2 antigen, are also highly desirable because of the co-involvement of these molecules in T cell activation, clonal expansion lymphokine (IL-2) secretion, and responsiveness to antigen. Given that both human B7.1 and B7.2 bind to 25 human CTLA-4 and CD28, it is probable that there is at least one common or homologous region (perhaps a shared conformational epitope or epitopes) to which macaque antibodies may potentially be raised.

30 The disclosed invention involves the use of an animal which is primed to produce a particular antibody. Animals which are useful for such a process include, but are not limited to, the following: mice, rats, guinea pigs, hamsters, monkeys, pigs, goats and rabbits.

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A preferred means of generating human antibodies using SCID mice is disclosed in commonly-owned, co-pending U.S. Patent application Serial No. 08/488,376.

The present inventors elected to immunize macaques against human B7.1 antigen using recombinant soluble B7.1 antigen produced in CHO cells and purified by affinity chromatography using a L307.4-sepharose affinity column. However, the particular source of human B7 antigen, human B7.1 antigen or human B7.2 antigen is not critical, provided that it is of sufficient purity to result in a specific antibody response to the particular administered B7 antigen and potentially to other B7 antigens.

The human B7 antigen, human B7.1 antigen (also called CD80) and human B7.2 antigen (also called CD86) genes have been cloned, and sequenced, and therefore may readily be manufactured by recombinant methods.

Preferably, the administered human B7 antigen, human B7.1 antigen and/or human B7.2 antigen will be administered in soluble form, e.g., by expression of a B7, B7.1 or B7.2 gene which has its transmembrane and cytoplasmic domains removed, thereby leaving only the extracellular portion, i.e., the extracellular superfamily V and C-like domains. (See, e.g., Grumet et al, Hum. Immunol., 40(3), p. 228-234, 1994, which teaches expression of a soluble form of human B7, which is incorporated by reference in its entirety herein).

The macaques will be immunized with the B7, B7.1 and/or B7.2 antigen, preferably a soluble form thereof, under conditions which result in the production of antibodies specific thereto. Preferably, the soluble human B7, B7.1 or B7.2 antigen will be administered in combination with an adjuvant, e.g., Complete Freund's Adjuvant (CFA), Alum, Saponin, or other known adjuvants,

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as well as combinations thereof. In general, this will require repeated immunization, e.g., by repeated injection, over several months. For example, administration of soluble B7.1 antigen was effected in 5 adjuvant, with booster immunizations, over a 3 to 4 month period, with resultant production of serum containing antibodies which bound human B7.1 antigen.

After immunization B cells are collected, e.g., by lymph node biopsies taken from the immunized animals and 10 B lymphocytes fused with KH6/B5 (mouse x human) heteromyeloma cells using polyethylene glycol. Methods for preparation of such heteromyelomas are known and may be found in U.S. Serial No. 08/379,072 by Newman et al, filed on January 25, 1995 and incorporated by reference 15 herein.

Heterohybridomas which secrete antibodies which bind human B7, B7.1 and/or B7.2 are then identified. This may be effected by known techniques. For example, this may be determined by ELISA or radioimmunoassay 20 using enzyme or radionucleotide labelled human B7, B7.1 and/or B7.2 antigen.

Cell lines which secrete antibodies having the desired specificity to human B7, B7.1 and/or B7.2 antigen are then subcloned to monoclonality.

25 In the present invention, the inventors screened purified antibodies for their ability to bind to soluble B7.1 antigen coated plates in an ELISA assay, antigen positive B cells, and CHO transfecomas which express human B7.1 antigen on their cell surface. In addition, 30 the antibodies were screened for their ability to block B cell/T cell interactions as measured by IL-2 production and tritiated thymidine uptake in a mixed lymphocyte reaction (MLR), with B7 binding being detected using ¹²⁵I-radiolabeled soluble B7.1 (SB7.1).

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Also, affinity purified antibodies from macaques were tested for their reactivity against CHO transfectants which expressed B7.1/Ig fusion proteins, and against CHO cells which produced human B7.2 antigen.

5 These results indicated that the B7.1 immune sera bound to the B7.2 transfecomas. Binding of antibodies to B7.2 antigen may be confirmed using soluble B7.2-Ig reagents. As discussed in the examples, this may be effected by producing and purifying B7.2-Ig from CHO 10 transfecomas in sufficient quantities to prepare a B7.2-Ig-sepharose affinity column. Those antibodies which cross-react with B7.2 will bind the B7.2-Ig-sepharose column.

Cell lines which express antibodies which 15 specifically bind to human B7 antigen, B7.1 antigen and/or B7.2 antigen are then used to clone variable domain sequences for the manufacture of primatized antibodies essentially as described in Newman et al, (1992), Id. and Newman et al, U.S. Serial No. 379,072, 20 filed January 25, 1995, both of which are incorporated by reference herein. Essentially, this entails extraction of RNA therefrom, conversion to cDNA, and amplification thereof by PCR using Ig specific primers. Suitable primers are described in Newman et al, 1992, 25 Id. and in U.S. Serial No. 379,072. (See, in particular, Figure 1 of U.S. Serial No. 379,072).

The cloned monkey variable genes are then inserted into an expression vector which contains human heavy and light chain constant region genes. Preferably, this is 30 effected using a proprietary expression vector of IDEC, Inc., referred to as NEOSPLA. This vector is shown in Figure 2 and contains the cytomegalovirus promoter/enhancer, the mouse beta globin major promoter, the SV40 origin of replication, the bovine growth

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hormone polyadenylation sequence, neomycin phosphotransferase exon 1 and exon 2, human immunoglobulin kappa or lambda constant region, the dihydrofolate reductase gene, the human immunoglobulin 5 gamma 1 or gamma 4 PE constant region and leader sequence. This vector has been found to result in very high level expression of primatized antibodies upon incorporation of monkey variable region genes, 10 transfection in CHO cells, followed by selection in G418 containing medium and methotrexate amplification.

For example, this expression system has been previously disclosed to result in primatized antibodies having high avidity ($K_d \leq 10^{-10}$ M) against CD4 and other human cell surface receptors. Moreover, the antibodies 15 have been found to exhibit the same affinity, specificity and functional activity as the original monkey antibody. This vector system is substantially disclosed in commonly assigned U.S. Serial No. 379,072, incorporated by reference herein as well as U.S. Serial 20 No. 08/149,099, filed on November 3, 1993, also incorporated by reference in its entirety herein. This system provides for high expression levels, i.e., > 30 pg/cell/day.

As discussed *infra*, the subject inventors have 25 selected four lead candidate monkey monoclonal antibodies which specifically bind the B7.1 antigen. These monkey monoclonal antibodies are referred to herein as 7B6, 16C10, 7C10 and 20C9.

As discussed in greater detail *infra*, these 30 antibodies were evaluated for their ability to block B cell/T cell interactions as measured by IL-2 production and tritiated thymidine uptake in a mixed lymphocyte reaction for T cell binding experiments for T cell binding, human buffy coat peripheral blood lymphocytes

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were cultured for 3-6 days in the presence of PHA stimulator. B7 binding was radioassayed using ^{125}I -radiolabeled soluble B7.1. The observed results indicate that all of these antibodies bind B7.1 antigen with high affinity and effectively block B cell/T cell interactions as evidenced by reduced IL-2 production and reduced proliferation of mixed lymphocyte cultures.

5. The properties of these particular monkey monoclonal antibodies are summarized below:

10. 1. Scatchard analysis showed that the apparent affinity constants (K_d) for the monkey antibodies binding to B7-Ig coated plates were approximated to be:

15. a: 7C10: $6.2 \times 10^{-9}\text{M}$
b: 16C10: $8.1 \times 10^{-9}\text{M}$
c: 7B6: $10.7 \times 10^{-9}\text{M}$
d: 20C9: $16.8 \times 10^{-9}\text{M}$

20. 2. The antibodies were tested in vitro in a mixed lymphocyte reaction assay (MLR). The MLR showed that all 4 anti-B7.1 antibodies inhibit IL-2 production to different extents as shown by the following IC_{50} values:

25. a: 7B6: $5.0 \mu\text{g/M}$
b: 16C10: $<0.1 \mu\text{g/M}$
c: 20C9: $2.0 \mu\text{g/M}$
d: 7C10: $5.0 \mu\text{g/M}$

30. 3. The monkey anti-B7.1 antibodies were tested for their ability to bind B7 on human peripheral blood lymphocytes (PBL). FACS analysis showed that all 4 monkey antibodies tested positive.

35. 4. Monkey antibodies 16C10, 7B6, 7C10 and 20C9 were tested for C1q binding by FACS analysis. Results showed 7C10 monkey Ig had strong human C1q binding after incubating with B7.1 CHO-

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transfected cells. 16C10 was positive, while 20C9 and 7B6 monkey antibodies were negative.

5. To select an animal model for path-tox studies, the monkey antibodies were tested with animal blood from different species. It was determined that the monkey anti-B7.1 antibodies cross-reacted with human, chimpanzee.

Based on these properties, it would appear that 10 three monkey monoclonal antibodies possess the most advantageous properties, 16C10, 7C10 and 20C9, with 16C10 and 7C10 being somewhat better than 20C9.

Using the techniques described *supra*, and in commonly assigned U.S. Serial No. 08/379,072, the 15 present inventors have cloned the variable domains of 7C10, 7B6 and 16C10, and provide the amino acid and nucleic acid sequences of primatized forms of the 7C10 light chain, 7C10 heavy chain, 7B6 light chain, 7B6 heavy chain, 16C10 light chain and 16C10 heavy chain. 20 These amino acid and nucleic acid sequences may be found in Figures 3a and 3b, 4a and 4b, and 5a and 5b. The DNA and amino acid sequence for the human gamma 1, gamma 4 constant domain may be found in 08/379,072.

As discussed *supra*, these primatized antibodies are 25 preferably expressed using the NEOSPLA expression vector shown in Figure 2 which is substantially described in commonly assigned 08/379,072 and 08/149,099, both of which applications are incorporated by reference herein.

As previously noted, the subject primatized 30 antibodies will preferably contain either the human immunoglobulin gamma 1 or gamma 4 constant region, with gamma 4 preferably mutated at two positions to create gamma 4 PE. The gamma 4 PE mutant contains two mutations, a glutamic acid in the CH2 region introduced

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to eliminate residual FCR binding, and a proline substitution in the hinge region, intended to enhance the stability of the heavy chain disulfide bond interaction. (See, Alegre et al, J. Immunol., 148, 5 3461-3468, (1992); and Angel et al, Mol. Immunol., 30, 105-158, (1993), both of which are incorporated by reference herein).

Whether the subject primatized antibodies contain the gamma 1, gamma 4 or gamma 4 PE constant region 10 largely depends on the particular disease target. Preferably, depleting and non-depleting primatized IgG1 and IgG4 antibodies are created and tested against specific disease targets.

Given the described binding and functional 15 properties of the subject monkey monoclonal antibodies, these anti-B7.1 monoclonal antibodies and primatized forms thereof should be well suited as therapeutic agents for blocking the B7:CD28 interaction thereby providing for immunosuppression. In particular, given 20 their high affinity to B7.1 antigen and ability to block B cell/T cell interactions as measured by IL-2 production and tritiated thymidine uptake in mixed lymphocyte culture as well as their ability to effectively inhibit antigen driven responses in donor 25 spleen cell cultures as shown by reduced antigen specific IgG responses, IL-2 production and cell proliferation, these monkey monoclonal antibodies and primatized forms thereof should function as effective immunosuppressants which modulate the B7:CD28 pathway. 30 This is significant for the treatment of many diseases wherein immunosuppression is therapeutically desirable, e.g., autoimmune diseases, to inhibit undesirable antigen specific IgG responses, and also for prevention of organ rejection and graft-versus-host disease.

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Essentially, the subject antibodies will be useful in treating any disease wherein suppression of the B7:CD28 pathway is therapeutically desirable.

Key therapeutic indications for the subject anti-B7.1 antibodies include, by way of example, autoimmune diseases such as idiopathic thrombocytopenia purpura (ITP), systemic lupus erythematosus (SLE), type 1 diabetes mellitus, multiple sclerosis, aplastic anemia, psoriasis, allergy, inflammatory bile disease and rheumatoid arthritis.

Another significant therapeutic indication of the subject anti-B7.1 antibodies is for prevention of graft-versus-host-disease (GVHD) during organ transplant and bone marrow transplant (BMT). The subject antibodies may be used to induce host tolerance to donor-specific alloantigens and thereby facilitate engraftment and reduce the incidence of graft rejection. It has been shown in a murine model of allogeneic cardiac transplantation that intravenous administration of CTLA4-Ig can result in immunosuppression or even induction of tolerance to alloantigen. (Lin et al, J. Exp. Med. 178:1801, 1993; Torka et al, Proc. Natl. Acad. Sci., USA, 89:11102, 1992). It is expected that the subject primatized anti-B7.1 antibodies will exhibit similar or greater activity.

Antibodies produced in the manner described above, or by equivalent techniques, can be purified by a combination of affinity and size exclusion chromatography for characterization in functional biological assays. These assays include determination of specificity and binding affinity as well as effector function associated with the expressed isotype, e.g., ADCC, or complement fixation. Such antibodies may be used as passive or active therapeutic agents against a

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number of human diseases, including B cell lymphoma, infectious diseases including viral diseases such as HIV/AIDS, autoimmune and inflammatory diseases, and transplantation. The antibodies can be used either in 5 their native form, or as part of an antibody/chelate, antibody/drug or antibody/toxin complex. Additionally, whole antibodies or antibody fragments (Fab₂, Fab, Fv) may be used as imaging reagents or as potential vaccines or immunogens in active immunotherapy for the generation 10 of anti-idiotypic responses.

The amount of antibody useful to produce a therapeutic effect can be determined by standard techniques well known to those of ordinary skill in the art. The antibodies will generally be provided by 15 standard technique within a pharmaceutically acceptable buffer, and may be administered by any desired route. Because of the efficacy of the presently claimed antibodies and their tolerance by humans it is possible to administer these antibodies repetitively in order to 20 combat various diseases or disease states within a human.

The anti-B7.1 antibodies (or fragments thereof) of this invention are useful for inducing 25 immunosuppression, i.e., inducing a suppression of a human's or animal's immune system. This invention therefore relates to a method of prophylactically or therapeutically inducing immunosuppression in a human or other animal in need thereof by administering an effective, non-toxic amount of such an antibody of this 30 invention to such human or other animal.

The ability of the compounds of this invention to induce immunosuppression has been demonstrated in standard tests used for this purpose, for example, a mixed lymphocyte reaction test or a test measuring

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inhibition of T-cell proliferation measured by thymidine uptake.

The fact that the antibodies of this invention have utility in inducing immunosuppression indicates that 5 they should be useful in the treatment or prevention of resistance to or rejection of transplanted organs or tissues (e.g., kidney, heart, lung, bone marrow, skin, cornea, etc.); the treatment or prevention of autoimmune, inflammatory, proliferative and 10 hyperproliferative diseases, and of cutaneous manifestations of immunologically mediated diseases (e.g., rheumatoid arthritis, lupus erythematosus, systemic lupus erythematosus, Hashimotos thyroiditis, multiple sclerosis, myasthenia gravis, type 1 diabetes, 15 uveitis, nephrotic syndrome, psoriasis, atopic dermatitis, contact dermatitis and further eczematous dermatitides, seborrheic dermatitis, Lichen planus, Pemphigus, bullous pemphigus, Epidermolysis bullosa, urticaria, angioedemas, vasculitides, erythema, 20 cutaneous eosinophilias, Alopecia areata, etc.); the treatment of reversible obstructive airways disease, intestinal inflammations and allergies (e.g., inflammatory bile disease, Coeliac disease, proctitis, eosinophilia gastroenteritis, mastocytosis, Crohn's 25 disease and ulcerative colitis), food-related allergies (e.g., migraine, rhinitis and eczema), and other types of allergies.

One skilled in the art would be able, by routine experimentation, to determine what an effective, non-toxic amount of antibody would be for the purpose of 30 inducing immunosuppression. Generally, however, an effective dosage will be in the range of about 0.05 to 100 milligrams per kilogram body weight per day.

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The antibodies (or fragments thereof) of this invention should also be useful for treating tumors in a mammal. More specifically, they should be useful for reducing tumor size, inhibiting tumor growth and/or 5 prolonging the survival time of tumor-bearing animals. Accordingly, this invention also relates to a method of treating tumors in a human or other animal by administering to such human or animal an effective, non-toxic amount of an antibody. One skilled in the art 10 would be able, by routine experimentation, to determine what an effective, non-toxic amount of anti-B7 antibody would be for the purpose of treating carcinogenic tumors. Generally, however, an effective dosage is expected to be in the range of about 0.05 to 100 15 milligrams per kilogram body weight per day.

The antibodies of the invention may be administered to a human or other animal in accordance with the aforementioned methods of treatment in an amount sufficient to produce such effect to a therapeutic or 20 prophylactic degree. Such antibodies of the invention can be administered to such human or other animal in a conventional dosage form prepared by combining the antibody of the invention with a conventional pharmaceutically acceptable carrier or diluent according 25 to known techniques. It will be recognized by one of skill in the art that the form and character of the pharmaceutically acceptable carrier or diluent is dictated by the amount of active ingredient with which it is to be combined, the route of administration and 30 other well-known variables.

The route of administration of the antibody (or fragment thereof) of the invention may be oral, parenteral, by inhalation or topical. The term parenteral as used herein includes intravenous,

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intraperitoneal, intramuscular, subcutaneous, rectal or vaginal administration. The subcutaneous and intramuscular forms of parenteral administration are generally preferred.

5 The daily parenteral and oral dosage regimens for employing compounds of the invention to prophylactically or therapeutically induce immunosuppression, or to therapeutically treat carcinogenic tumors will generally be in the range of about 0.05 to 100, but preferably
10 about 0.5 to 10, milligrams per kilogram body weight per day.

The antibodies of the invention may also be administered by inhalation. By "inhalation" is meant intranasal and oral inhalation administration.

15 Appropriate dosage forms for such administration, such as an aerosol formulation or a metered dose inhaler, may be prepared by conventional techniques. The preferred dosage amount of a compound of the invention to be employed is generally within the range of about 10 to
20 100 milligrams.

The antibodies of the invention may also be administered topically. By topical administration is meant non-systemic administration and includes the application of an antibody (or fragment thereof) compound of the invention externally to the epidermis, to the buccal cavity and instillation of such an antibody into the ear, eye and nose, and where it does not significantly enter the blood stream. By systemic administration is meant oral, intravenous, intraperitoneal and intramuscular administration. The amount of an antibody required for therapeutic or prophylactic effect will, of course, vary with the antibody chosen, the nature and severity of the condition being treated and the animal undergoing

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treatment, and is ultimately at the discretion of the physician. A suitable topical dose of an antibody of the invention will generally be within the range of about 1 to 100 milligrams per kilogram body weight 5 daily.

Formulations

While it is possible for an antibody or fragment thereof to be administered alone, it is preferable to present it as a pharmaceutical formulation. The active 10 ingredient may comprise, for topical administration, from 0.001% to 10% w/w, e.g., from 1% to 2% by weight of the formulation, although it may comprise as much as 10% w/w but preferably not in excess of 5% w/w and more preferably from 0.1% to 1% w/w of the formulation.

15 The topical formulations of the present invention, comprise an active ingredient together with one or more acceptable carrier(s) therefor and optionally any other therapeutic ingredients(s). The carrier(s) must be "acceptable" in the sense of being compatible with the 20 other ingredients of the formulation and not deleterious to the recipient thereof.

Formulations suitable for topical administration include liquid or semi-liquid preparations suitable for penetration through the skin to the site of where 25 treatment is required, such as liniments, lotions, creams, ointments or pastes, and drops suitable for administration to the eye, ear or nose.

Drops according to the present invention may 30 comprise sterile aqueous or oily solutions or suspensions and may be prepared by dissolving the active ingredient in a suitable aqueous solution of a bactericidal and/or fungicidal agent and/or any other suitable preservative, and preferably including a

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surface active agent. The resulting solution may then be clarified by filtration, transferred to a suitable container which is then sealed and sterilized by autoclaving or maintaining at 90°-100°C for half an 5 hour. Alternatively, the solution may be sterilized by filtration and transferred to the container by an aseptic technique. Examples of bactericidal and fungicidal agents suitable for inclusion in the drops are phenylmercuric nitrate or acetate (0.002%), 10 benzalkonium chloride (0.01%) and chlorhexidine acetate (0.01%). Suitable solvents for the preparation of an oily solution include glycerol, diluted alcohol and propylene glycol.

Lotions according to the present invention include 15 those suitable for application to the skin or eye. An eye lotion may comprise a sterile aqueous solution optionally containing a bactericide and may be prepared by methods similar to those for the preparation of drops. Lotions or liniments for application to the skin 20 may also include an agent to hasten drying and to cool the skin, such as an alcohol or acetone, and/or a moisturizer such as glycerol or an oil such as castor oil or arachis oil.

Creams, ointments or pastes according to the 25 present invention are semi-solid formulations of the active ingredient for external application. They may be made by mixing the active ingredient in finely-divided or powdered form, alone or in solution or suspension in an aqueous or non-aqueous fluid, with the aid of 30 suitable machinery, with a greasy or non-greasy basis. The basis may comprise hydrocarbons such as hard, soft or liquid paraffin, glycerol, beeswax, a metallic soap; a mucilage; an oil of natural origin such as almond, corn, arachis, castor or olive oil; wool fat or its

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derivatives, or a fatty acid such as stearic or oleic acid together with an alcohol such as propylene glycol or macrogols. The formulation may incorporate any suitable surface active agent such as an anionic, 5 cationic or non-ionic surface active such as sorbitan esters or polyoxyethylene derivatives thereof. Suspending agents such as natural gums, cellulose derivatives or inorganic materials such as silicaceous silicas, and other ingredients such as lanolin, may also 10 be included.

The subject anti-B7.1 antibodies or fragments thereof may also be administered in combination with other moieties which modulate the B7:CD28 pathway. Such moieties include, by way of example, cytokines such as 15 IL-7 and IL-10, CTLA4-Ig, soluble CTLA-4 and anti-CD28 antibodies and fragments thereof. Also, the subject antibodies may be administered in combination with other immunosuppressants. Such immunosuppressants include small molecules such as cyclosporin A (CSA) and FK506; 20 monoclonal antibodies such as anti-tumor necrosis factor a (anti-TNF α), anti-CD54, anti-CD11, anti-CD11a, and anti-IL-1; and, other soluble receptors such as rTNF α and rIL-1.

It will be recognized by one of skill in the art 25 that the optimal quantity and spacing of individual dosages of an antibody or fragment thereof of the invention will be determined by the nature and extent of the condition being treated, the form, route and site of administration, and the particular animal being treated, 30 and that such optimums can be determined by conventional techniques. It will also be appreciated by one of skill in the art that the optimal course of treatment, i.e., the number of doses of an antibody or fragment thereof of the invention given per day for a defined number of

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days, can be ascertained by those skilled in the art using conventional course of treatment determination tests.

Without further elaboration, it is believed that 5 one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following formulations are, therefore, to be construed as merely illustrative 10 embodiments and not a limitation of the scope of the present invention in any way.

Capsule Composition

A pharmaceutical composition of this invention in the form of a capsule is prepared by filling a standard two-piece hard gelatin capsule with 50 mg. of an 15 antibody or fragment thereof of the invention, in powdered form, 100 mg. of lactose, 32 mg. of talc and 8 mg. of magnesium stearate.

Injectable Parenteral Composition

A pharmaceutical composition of this invention in a 20 form suitable for administration by injection is prepared by stirring 1.5% by weight of an antibody or fragment thereof of the invention in 10% by volume propylene glycol and water. The solution is sterilized by filtration.

25 Ointment Composition

Antibody or fragment thereof of the invention 1.0 g.

White soft paraffin to 100.0 g.

The antibody or fragment thereof of the invention 30 is dispersed in a small volume of the vehicle to produce

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a smooth, homogeneous product. Collapsible metal tubes are then filled with the dispersion.

Topical Cream Composition

Antibody or fragment thereof of the invention

5 1.0 g.

Polawax GP 200 20.0 g.

Lanolin Anhydrous 2.0 g.

White Beeswax 2.5 g.

Methyl hydroxybenzoate 0.1 g.

10 Distilled Water to 100.0 g.

The polawax, beeswax and lanolin are heated together at 60°C. A solution of methyl hydroxybenzoate is added and homogenization is achieved using high speed stirring. The temperature is then allowed to fall to 15 50°C. The antibody or fragment thereof of the invention is then added and dispersed throughout, and the composition is allowed to cool with slow speed stirring.

Topical Lotion Composition

Antibody or fragment thereof of the invention

20 1.0 g.

Sorbitan Monolaurate 0.6 g.

Polysorbate 20 0.6 g.

Cetostearyl Alcohol 1.2 g.

Glycerin 6.0 g.

25 Methyl Hydroxybenzoate 0.2 g.

Purified Water B.P. to 100-00 ml. (B.P. = British Pharmacopeia)

The methyl hydroxybenzoate and glycerin are dissolved in 70 ml. of the water at 75°C. The sorbitan monolaurate, polysorbate 20 and cetostearyl alcohol are melted together at 75°C and added to the aqueous

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solution. The resulting emulsion is homogenized, allowed to cool with continuous stirring and the antibody or fragment thereof of the invention is added as a suspension in the remaining water. The whole
5 suspension is stirred until homogenized.

Eye Drop Composition

Antibody or fragment thereof of the invention
0.5 g.

10 Methyl Hydroxybenzoate 0.01 g.
Propyl Hydroxybenzoate 0.04 g.
Purified Water B.P. to 100-00 ml.

15 The methyl and propyl hydroxybenzoates are dissolved in 70 ml. purified water at 75°C and the resulting solution is allowed to cool. The antibody or fragment thereof of the invention is then added, and the solution is sterilized by filtration through a membrane filter (0.022 μ m pore size), and packed aseptically into suitable sterile containers.

Composition for Administration by Inhalation

20 For an aerosol container with a capacity of 15-20 ml: mix 10 mg. of an antibody or fragment thereof of the invention with 0.2-0.5% of a lubricating agent, such as polysorbate 85 or oleic acid, and disperse such mixture in a propellant, such as freon, preferably in a
25 combination of (1,2 dichlorotetrafluoroethane) and difluorochloro-methane and put into an appropriate aerosol container adapted for either intranasal or oral inhalation administration.

Composition for Administration by Inhalation

30 For an aerosol container with a capacity of 15-20 ml: dissolve 10 mg. of an antibody or fragment thereof

of the invention in ethanol (6-8 ml.), add 0.1-0.2% of a lubricating agent, such as polysorbate 85 or oleic acid; and disperse such in a propellant, such as freon, preferably in combination of (1.2 dichlorotetrafluoroethane) and difluorochloromethane, and put into an appropriate aerosol container adapted for either intranasal or oral inhalation administration.

5 The antibodies and pharmaceutical compositions of the invention are particularly useful for parenteral administration, i.e., subcutaneously, intramuscularly or intravenously. The compositions for parenteral administration will commonly comprise a solution of an antibody or fragment thereof of the invention or a cocktail thereof dissolved in an acceptable carrier, 10 preferably an aqueous carrier. A variety of aqueous carriers may be employed, e.g., water, buffered water, 0.4% saline, 0.3% glycine, and the like. These 15 solutions are sterile and generally free of particulate matter. These solutions may be sterilized by 20 conventional, well-known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate 25 physiological conditions such as pH adjusting and buffering agents, etc. The concentration of the antibody or fragment thereof of the invention in such pharmaceutical formulation can vary widely, i.e., from less than about 0.5%, usually at or at least about 1% to as much as 15 or 20% by weight, and will be selected 30 primarily based on fluid volumes, viscosities, etc., according to the particular mode of administration selected.

Thus, a pharmaceutical composition of the invention for intramuscular injection could be prepared to contain 1 Ml sterile buffered water, and 50 mg. of an antibody

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or fragment thereof of the invention. Similarly, a pharmaceutical composition of the invention for intravenous infusion could be made up to contain 250 ml. of sterile Ringer's solution, and 150 mg. of an antibody 5 or fragment thereof of the invention. Actual methods for preparing parenterally administrable compositions are well known or will be apparent to those skilled in the art, and are described in more detail in, for example, Remington's Pharmaceutical Science, 15th ed., 10 Mack Publishing Company, Easton, Pennsylvania, hereby incorporated by reference herein.

The antibodies (or fragments thereof) of the invention can be lyophilized for storage and reconstituted in a suitable carrier prior to use. This 15 technique has been shown to be effective with conventional immune globulins and art-known lyophilization and reconstitution techniques can be employed.

Depending on the intended result, the 20 pharmaceutical composition of the invention can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from a disease, in an amount sufficient to cure or at least 25 partially arrest the disease and its complications. In prophylactic applications, compositions containing the present antibodies or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance.

30 Single or multiple administrations of the pharmaceutical compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical composition of the invention should provide a quantity of the

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altered antibodies (or fragments thereof) of the invention sufficient to effectively treat the patient.

It should also be noted that the antibodies of this invention may be used for the design and synthesis of either peptide or non-peptide compounds (mimetics) which would be useful in the same therapy as the antibody. See, e.g., Saragovi et al., Science, 253, 792-795 (1991).

To further illustrate the invention, the following examples are provided. These examples are not intended, nor are they to be construed, as further limiting the invention.

Example 1

Recombinant immunoglobulin libraries displayed on the surface of filamentous phage were first described by McCafferty et al, Nature, 348:552-554, 1990 and Barbas et al, Proc. Natl. Acad. Sci., USA 88:7978-7982, 1991. Using this technology, high affinity antibodies have been isolated from immune human recombinant libraries (Barbas et al, Proc. Natl. Acad. Sci., USA 89:10164-10168, 1992). Although the phage display concept used is substantially similar to that described by Barbas, 1991, Id. the technique has been modified by the substitution of a unique vector for monkey libraries to reduce the possibility of recombination and improve stability. This vector, pMS, Figure 1 contains a single lac promoter/operator for efficient transcription and translation of polycistronic heavy and light chain monkey DNA. This vector contains two different leader sequences, the omp A (Movva et al, J. Biol. Chem., 255: 27-29, (1980), for the light chain and the pel B (Lei, J. Bact., 4379-109:4383 (1987) for the heavy chain Fd. Both leader sequences are translated into hydrophobic

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signal peptides that direct the secretion of the heavy and light chain cloned products into the periplasmic space. In the oxidative environment of the periplasm, the two chains fold and disulfide bonds form to create 5 stable Fab fragments. We derived the backbone of the vector from the phagemid bluescript. (Stratagene, La Jolla, CA). It contains the gene for the enzyme beta-lactamase that confers ampicillin (carbenicillin) resistance to bacteria that harbor pMS DNA. We also 10 derived, from bluescript, the origin of replication of the multicopy plasmid ColE1 and the origin of replication of the filamentous bacteriophage f1. The origin of replication of phage f1 (the so-called intragenic region), signals the initiation of synthesis 15 of single stranded pMS DNA, the initiation of capsid formation and the termination of RNA synthesis by viral enzymes. The replication and assembly of pMS DNA strands into phage particles requires viral proteins that must be provided by a helper phage. We have used 20 helper phage VCSM13 which is particularly suited for this, since it also contains a gene coding for kanamycin resistance. Bacteria infected with VCSM13 and pMS can be selected by adding both kanamycin and carbenicillin to the growth medium. The bacteria will ultimately 25 produce filamentous phage particles containing either pMS or VCSM13 genomes. Packaging of the helper phage is less efficient than that of pMS, resulting in a mixed phage population that contains predominately recombinant pMS phages. The ends of the phage pick up minor coat 30 proteins specific to each end. Of particular interest here is the gene III product which is present in three to five copies at one end of the phage. The gene III product is 406 amino acid residues and is required for phage infection of *E. coli* via the F pili. The first

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two domains of the heavy chain, the variable and the CH1 domain, are fused to the carboxy-terminal half of the gene III protein. This recombinant pili protein, directed by the pel B leader, is secreted to the

5 periplasm where it accumulates and forms disulfide bonds with the light chain before it is incorporated in the coat of the phage. Also, another vector contains a FLAG sequence engineered downstream of the gene III. The FLAG is an 8 amino acid peptide expressed at the carboxy terminal of the Fd protein. We are using commercially available monoclonal anti-FLAG M2 for both purification and detection of phage Fab by ELISA (Brizzard, Bio Techniques, 16(4):730-731, (1994)).

After constructing the vector pMS, we tested its ability to produce phage bound Fab using control antibody genes. We cloned an anti-tetanus toxoid antibody, (obtained from Dr. Carlos Barbas), into pMS and transformed XL1-blue. We co-infected our cells with VCSM13 and generated phage displaying the anti-tetanus toxoid antibody. We performed efficiency experiments where anti-tetanus toxoid phage were combined with phage bearing an irrelevant antibody at 1:100,000. We performed three rounds of panning by applying 50 μ l of the mixed phage to antigen (tetanus toxoid) coated

15 20 25 polystyrene wells. Non-adherent phage were washed off and the adherent phage were eluted with acid. The eluted phage were used to infect a fresh aliquot of XL1-Blue bacteria and helper phage was added. After overnight amplification, phage were prepared and again panned on antigen coated plates. After three rounds of

30 panning, we were able to show that we had successfully enriched for the anti-tetanus toxoid phage. The success of this technology also depends on the ability to prepare soluble Fabs for characterization of the final

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panned product. This was achieved by excising gene III from the pMS DNA using the restriction enzyme Nhe I followed by re-ligation. After the gene III was excised, the Fab was no longer displayed on the phage 5 surface but accumulated in the piroplasmic space. Lysates were prepared from bacteria expressing soluble Fab and tested for antigen specificity using an ELISA. High levels of soluble Fab were detected.

In order to adapt phage display technology for use 10 with macaque libraries, we developed specific primers for PCR amplifying monkey immunoglobulin genes. These were based on macaque sequences we obtained while developing the PRIMATIZED® antibody technology (See, 08/379,072, incorporated by reference herein) and 15 databases containing human sequences. (Kabat et al, (1991), "Sequences of Proteins of Immunological Interest," U.S. Dept. of Health and Human Services, National Institute of Health).

We developed three sets of primers to cover 20 amplification of the macaque repertoire. Our first set of primers was designed for amplification of the heavy chain VH and CH1 (Fd) domains. It consisted of a 3' CH1 domain primer and six 5' VH family specific primers that bind in the framework 1 region. Our second set of 25 primers, for amplifying the whole lambda chain, covers the many lambda chain subgroups. It consists of a 3' primer and three 5' degenerate primers that bind in the VL framework 1 region. Our third set of primers was designed for amplification of the kappa chain subgroups. 30 It consists of one 3' primer and five VK framework 1 primers. Using each of these sets, PCR parameters were optimized to obtain strong enough signals from each primer pair so that ample material was available for cloning of the library. We recently created macaque

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combinatorial libraries in our pMS vector using these optimized PCR conditions. Bone marrow biopsies were taken from CD4 immune monkeys as the source of immunoglobulin RNA. The libraries contained 5 approximately 10^6 members and are currently being panned or specific binders on antigen coated wells.

Example 2

Development of B7/CTLA-4 Reagents

We have generated a number of reagents for the 10 purpose of immunizing monkeys, developing binding and functional assays *in vitro*, screening heterohybridomas and panning phage libraries. Table 1 lists each reagent and its intended purpose. In the case of B7.1, RNA was extracted from SB cells and converted to cDNA using 15 reverse transcriptase. The first strand cDNA was PCR amplified using B7.1 specific primers and cloned into IDEC's NEOSPLA mammalian expression vectors. CHO cells were transfected with B7.1 NEOSPLA DNA and clones expressing membrane associated B7.1 were identified. 20 The B7.1 fusion protein was generated similarly, except that the PCR amplified B7.1 gene was cloned into a NEOSPLA cassette vector containing the human CH2 and CH3 immunoglobulin genes. CHO cells were transformed with the B7.1/Ig NEOSPLA DNA and stable clones secreting 25 B7.1/Ig fusion protein were amplified. In general, the B7.2 and CTLA4 reagents were generated in the same manner, except that for B7.2 the RNA was isolated from human spleen cells that had been stimulated 24 hours with anti-Ig and IL-4, and for the CTLA4 constructs the 30 gene source was PHA activated human T cells.

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Table 1

Reagent	Purpose	CHO Expression
5	Soluble B7.1	Immunization, immunoassays
	B7.1 Transfectant	Screening, ELISA
	B7.1/Ig Fusion Protein	Inhibition studies, panning
	B7.2 Transfectant	Screening, ELISA
	B7.2/Ig Fusion Protein	Inhibition studies, panning
	CTLA4 Transfectant	Inhibition studies
	CTLA4/Ig	Inhibition studies

10 The availability of these reagents, together with monoclonal antibodies to B7.1 (L3074) (Becton Dickinson, 1994) and B7.2 (Fun-1 (Engel et al, Blood, 84, 1402-1407, (1994) and purified goat and rabbit antisera, specifically developed to detect monkey Fab fragments, 15 facilitates identification of antibodies having the desired properties.

Example 3

Generation of a Phage Display Library

20 Recombinant phage display libraries are generated from B7.1 and B7.2 immune monkeys. Lymph node and bone marrow biopsies are performed 7-12 days after immunization to harvest RNA rich B cells and plasma cells. RNA is isolated from the lymphocytes using the method described by Chomczynski Anal. Biochem., 162(1), 156-159, (1987). RNA is converted to cDNA using an oligo dT primer and reverse transcriptase. The first strand cDNA is divided into aliquots and PCR amplified using the sets of kappa, lambda, and heavy chain Fd region primers described earlier and either Pfu 25 polymerase (Stratagene, San Diego) or Taq polymerase (Promega, Madison). The heavy chain PCR amplified products are pooled, cut with Xho VSpe I restriction enzymes and cloned into the vector pMS. Subsequently, 30

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the light chain PCR products are pooled, cut with Sac I/Xba I restriction enzymes, and cloned to create the recombinant library. XLI-Blue *E. coli* is transformed with the library DNA and super-infected with 5 VCSM13 to produce the phage displaying antibodies. The library is panned four rounds on polystyrene wells coated with B7.1 or B7.2 antigen. Individual phage clones from each round of panning are analyzed. The pMS vector DNA is isolated and the gene III excised. 10 Soluble Fab fragments are generated and tested in ELISA for binding to B7.1 and B7.2.

Example 4

Characterization of Phage Fab Fragments

The monkey phage Fab fragments are characterized 15 for their specificity and the ability to block B7.1-Ig and B7.2-Ig binding to CTLA-4-Ig or CTLA-4 transfected cells. Phage fragments are also characterized for cross-reactivity after first panning for 4 rounds on the B7 species used for immunization in order to select for 20 high affinity fragments. Fab fragments identified from four rounds of panning either on B7.1 or B7.2 antigen coated surfaces are scaled up by infection and grown in 24 hour fermentation cultures of *E. coli*. Fragments are purified by Kodak FLAG binding to a anti-FLAG affinity 25 column. Purified phage Fabs are tested for affinity by an ELISA based direct binding modified Scatchard analysis (Katch et al, J. Chem. BioEng., 76:451-454, (1993)) using Goat anti-monkey Fab antibodies or anti-FLAG MAb conjugated with horseradish peroxidase. The 30 anti-monkey Fab reagents will be absorbed against human heavy chain constant region Ig to remove any cross-reactivity to B7-Ig. Kd values are calculated for each

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fragment after measurements of direct binding to B7.1-Ig or B7.2-Ig coated plates.

Example 5

Phage Fab Fragment Blocking of CTLA-4/B7 Binding

5 Fab fragments most effectively blocking the binding of B7-Ig at the lowest concentrations are selected as lead candidates. Selections are made by competing off ^{125}I -B7-Ig binding to CTLA-4-Ig or CTLA-4 transfected cells. Additional selection criteria include, blocking 10 of mixed lymphocyte reaction (MLR), as measured by inhibiting ^3H -thymidine uptake in responder cells (Azuma et al, J. Exp. Med., 177:845-850,; Azuma et al, Nature, 301:76-79, (1993)) and direct analysis of IL-2 production using IL-2 assay kits. The three or four 15 candidates which are most effective in inhibiting of MLR and CTLA-4 binding assays are chosen for cloning into the above-described mammalian expression vector for transfection into CHO cells and expression of chimeric monkey/human antibodies.

20

Example 6

Generation of Monkey Heterohybridomas

Monkey heterohybridomas secreting monoclonal antibodies are generated from existing immunized animals whose sera tested positive for B7.1 and/or B7.2. Lymph 25 node biopsies are taken from animals positive to either, or both, antigens. The method of hybridoma production is similar to the established method used for the generation of monkey anti-CD4 antibodies (Newman, 1992(Id.)). Monkeys with high serum titers will have 30 sections of inguinal lymph nodes removed under

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anesthesia. Lymphocytes are washed from the tissue and fused with KH6/B5 heteromyeloma cells (Carrol et al, J. Immunol. Meth., 89:61-72, (1986)) using polyethylene glycol (PEG). Hybridomas are selected on H.A.T. media 5 and stabilized by repeated subcloning in 96 well plates.

Monkey monoclonal antibodies specific for B7.1 antigen are screened for cross-reactivity to B7.2. Monkey anti-B7 antibodies will be characterized for blocking of B7/CTLA-4 binding using the ^{125}I -B7-Ig 10 binding assay. Inhibition of MLR by ^3H -Thymidine uptake and direct measurement of IL-2 production is used to select three candidates. Two candidates will be brought forward in Phase II studies and expressed in CHO cells while repeating all functional studies. For the 15 purposes of developing an animal model for *in vivo* pharmacology, anti-B7 antibodies will be tested on cells of several animal species. The establishment of an animal model will allow preclinical studies to be carried out for the selected clinical indication.

20

Example 7

As discussed *supra*, using the above heterohybridoma methods, 4 lead monkey anti-B7.1 antibodies have been identified: 16C10, 7B6, 7C10 and 20C9. These antibodies were characterized as follows:

25

Scatchard analysis showed that the apparent affinity constants (K_d) for the monkey antibodies binding to B7-Ig coated plates were approximated to be:

30

a: 7C10: $6.2 \times 10^{-9}\text{M}$
b: 16C10: $8.1 \times 10^{-9}\text{M}$
c: 7B6: $10.7 \times 10^{-9}\text{M}$
d: 20C9: $16.8 \times 10^{-9}\text{M}$

The antibodies were tested *in vitro* in a mixed lymphocyte reaction assay (MLR). The MLR showed

-57-

that all 4 anti-B7.1 antibodies inhibit IL-2 production to different extents:

5 a: 7B6: 5.0 μ g/Ml
 b: 16C10: 0.1 μ g/Ml
 c: 20C9: 2.0 μ g/Ml
 d: 7C10: 5.0 μ g/Ml

10 The monkey anti-B7.1 antibodies were tested for their ability to bind B7 on human peripheral blood lymphocytes (PBL). FACS analysis showed that all 4 monkey antibodies tested positive.

15 Monkey antibodies 16C10, 7B6, 7C10 and 20C9 were tested for C1q binding by FACS analysis. Results showed 7C10 monkey Ig had strong human C1q binding after incubating with B7.1 CHO-transfected cells.

16C10 was also positive, while 20C9 and 7B6 monkey antibodies were negative.

Example 8

20 Using the primate antibody methodology incorporated by reference to commonly assigned U.S. Serial No. 08/379,072, and using the NEOSPLA vector system shown in Figure 2, the heavy and light variable domains of 7C10, 7B6 and 16C10 were cloned and primate forms thereof have been synthesized in CHO cells using the NEOSPLA vector system. The amino acid 25 and nucleic acid sequences for the primate 7C10 light and heavy chain, 7B6 light and heavy chain, and 16C10 light and heavy chain are respectively shown in Figures 3a, 3b, 4a, 4b, 5a and 5b.

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Example 9

Confirming experiments on the non-cross-reactivity of the CTLA-4 and PRIMATIZED® antibody binding sites on B7.1.

5 In competitive binding assays using biotinylated CTLA-4Ig (Figure 6), unlabeled primatized 16C10 (i.e., P16C10) was unable to block CTLA-4Ig binding to B7.1 transfected CHO cells. It can be seen that unlabeled CTLA-4Ig and unlabeled B7.1 effectively compete under 10 these conditions.

In a second experiment using Biotinylated P16C10, the same conclusions can be made. In the experiment shown in Figure 7, binding of P16C10-Biotin is inhibited by both unlabeled P16C10 and B7.1Ig, but not by CTLA-15 4Ig. Although CTLA-4Ig is reported to be as much as 4-10 fold higher in affinity ($K_d=0.4$ nM; Morton et al., J. Immunol. 156:1047-1054 (1996)), there is no significant inhibition of P16C10 binding even at CTLA-4Ig concentrations as high as 100 fold excess.

20 Similar results were obtained using the primatized antibody 7C10 (P7C10) when it was substituted for P16C10 in the experiments (data not provided).

Example 10

25 Comparing the ability of L307.4 and BB-1 mouse antibodies to bind to B7 CHO cells in the presence of CTLA-4Ig.

The binding of L307.4 and BB-1 murine anti-B7 antibody in the presence of CTLA-4Ig was studied in order to determine whether the mouse antibody binding 30 sites overlapped with the CTLA-4 binding site. Competition assay experiments using P16C10-Biotin, L307.4-Biotin and CTLA-4Ig-Biotin were done to insure that affinity differences did not prevent detection of

-59-

competitive binding. The results are shown in Figures 8 and 9.

The results of Figure 8 confirm earlier studies that the mouse antibody BB-1 does not compete with P16C10. These results also show that there is some cross-reactivity to L307.4 of approximately 50%. The results of Figure 8 confirm that BB-1 and L307.4 both compete with each other and that BB-1 completely blocks binding of CTLA-4Ig-Biotin to B7.1 transfected CHO cells. BB-1 does not significantly affect P16C10 binding to B7.1 positive CHO cells.

The results shown in Figure 9 indicate better than 50% competition when CTLA-4Ig-Biotin is used in the binding experiment. Figure 9 shows that CTLA-4Ig-Biotin is effectively blocked by all B7.1 inhibitors except P16C10, therefore P16C10 recognizes a unique binding determinate on B7.1 which allows the normal CTLA-4 ligand binding in the generation of negative signals. Earlier functional studies (data not shown) suggest a weakened ability of L307.4 to block IL-2 production in allogeneic MLR, which correlates with the hypothesis that it may interfere with CTLA-4 negative signaling. It is not clear how many of the other murine antibodies reported in the literature give complete inhibition of CTLA-4 binding; however, this issue may be important in defining the true functional mechanisms of B7.1 and B7.2 specific antibodies.

These results confirm earlier studies using B7-Ig in competition with P16C10-Biotin for binding to B7.1 transfected CHO cells. The studies also confirm earlier observations of no inhibition of the P16C10 by CTLA-4Ig. These results are highly suggestive that the primate antibodies are specific for a unique B7.1 epitope independent of the CTLA-4 binding site which interacts

-60-

primarily with CD28. This type of interaction would provide a benefit, since it has the ability to block binding of B7.1 to CD28 receptors while still allowing the negative signaling function of CTLA-4 to occur
5 uninhibited. This perceived interaction may lead to a down regulation of the overall T cell activation response regardless of the predominance of either Th1 or Th2 phenotypes.

Similar results were obtained using P7C10 when it
10 was substituted for P16C10 in the experiments (data not provided).

Example 11

Experiment demonstrating the ability of P16C10 to bind and block B7.1 interactions with CD28 receptor.

15 An experiment to determine if P16C10 binding of B7.1 can block the interaction of B7.1 with CD28 was attempted by radiolabeling B7.1Ig with ^{125}I , followed by competitive binding to CD28 positive non-activated peripheral blood T lymphocytes. The results shown in
20 Figure 10 demonstrate that the radiolabeled B7.1Ig binds specifically to the T cells, as confirmed by inhibition with unlabeled B7.1Ig. The results also show that CTLA-4Ig, anti-CD28 and P16C10 are all capable of blocking this interaction. The results further confirm that
25 P16C10 blocks binding of the CD28/B7 interaction with an IC₅₀ of < 1 ug/mL.

The above results were obtained under conditions where no membrane associated CTLA-4 was expressed (Linsley et al., J. Exp. Med. 173:721-730 (1991)) and
30 confirmed by blocking with the anti-CD28 antibody.

Similar results were obtained using P7C10 when it was substituted for P16C10 in the experiments (data not provided).

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It is expected that these primatized antibodies, given their probable low antigenicity and human effector function, will be well suited as therapeutics. In fact, it has recently been shown that primatized 16C10

5 exhibits human Clq binding.

Those skilled in the art will recognize or be able to ascertain using no more than routine experimentation many equivalents to the specific embodiments of the invention described herein. Such equivalents are

10 intended to be embraced by the following claims.

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What Is Claimed Is:

1. A monoclonal antibody which specifically binds to B7.1 antigen (CD80) or B7.2 antigen (CD86), and which antibody inhibits the binding of said B7.1 or B7.2 antigen to CD28.
2. The monoclonal antibody of claim 1 wherein said antibody specifically binds to B7.1 antigen (CD80).
3. The monoclonal antibody of claim 2 wherein said antibody does not inhibit the binding of B7.1 antigen to CTLA-4.
4. The monoclonal antibody of claim 1 wherein said antibody specifically binds to B7.2 antigen (CD86).
5. The monoclonal antibody of claim 4 wherein said antibody does not inhibit the binding of B7.2 antigen to CTLA-4.
6. The monoclonal antibody of claim 1 which inhibits the production of IL-2 by T cells.
7. The monoclonal antibody of claim 2 which selectively inhibits the interaction of B and T cells via the CD28/B7.1 pathway.
8. The monoclonal antibody of claim 4 which selectively inhibits the interaction of B and T cells via the CD28/B7.2 pathway.
9. The monoclonal antibody of claim 1 which is capable of inhibiting *in vitro* the production of IL-2 by T lymphocytes.

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10. The monoclonal antibody of claim 9 wherein said antibody is capable of inhibiting IL-2 production when added to a T lymphocyte containing culture at a concentration of at least 10 μ g/ml.

11. A monoclonal antibody which binds to the same epitope on B7.1 as 16C10 or 7C10, or which monoclonal antibody inhibits the interaction of 16C10 or 7C10 with B7.1.

12. The monoclonal antibody of claim 1 which is a primatized antibody.

13. The monoclonal antibody of claim 1 which is a human, chimeric mouse/human, or humanized antibody.

14. The monoclonal antibody of claim 1 wherein said B7.1 is human B7.1.

15. The monoclonal antibody of claim 1 wherein said B7.2 is human B7.2.

16. A method of treating a disease involving T cell/B cell interactions comprising administering an amount of a monoclonal antibody according to claim 2 sufficient to inhibit the binding of B cells and T cells via the B7.1/CD28 pathway.

17.. A method of treating a disease involving T cell/B cell interactions comprising administering an amount of a monoclonal antibody according to claim 4 sufficient to inhibit the binding of B cells and T cells via the B7.2/CD28 pathway.

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18. The method of claim 16 wherein said disease is an autoimmune disorder.

19. The method of claim 17 wherein said disease is an autoimmune disorder.

20. The method of claim 16 wherein said disease is selected from the group consisting of idiopathic thrombocytopenia purpura, systemic lupus erythematosus, type 1 diabetes mellitus, rheumatoid arthritis, psoriasis, aplastic anemia, inflammatory bile disease, allergy and multiple sclerosis.

21. The method of claim 17 wherein said disease is selected from the group consisting of idiopathic thrombocytopenia purpura, systemic lupus erythematosus, type 1 diabetes mellitus, rheumatoid arthritis, psoriasis, aplastic anemia, inflammatory bile disease, allergy and multiple sclerosis.

22. The method of claim 16 wherein said disease is graft-versus-host disease.

23. The method of claim 17 wherein said disease is graft-versus-host disease.

24. The method of claim 16 wherein said disease is selected from the group consisting of B cell lymphoma, infectious diseases, and inflammatory diseases.

25. The method of claim 17 wherein said disease is selected from the group consisting of B cell lymphoma, infectious diseases, and inflammatory diseases.

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26. A pharmaceutical composition suitable for treatment of a disease treatable by inhibition of B7:CD28 binding which comprises an antibody according to claim 1.

27. The method of claim 16 wherein the antibody is administered in combination with other recombinant protein or small molecule immunosuppressants.

28. The method of claim 17 wherein the antibody is administered in combination with other recombinant protein or small molecule immunosuppressants.

FIGURE 1

Figure 1: pMS Phage Display Vector. Pro = Lac Z promoter. L1 = Omp A leader. L2 = Pet B leader. Light = light chain. Heavy = heavy chain.

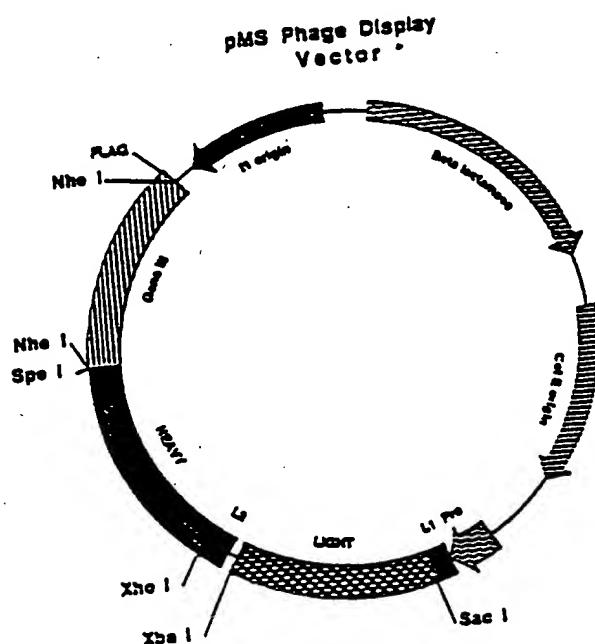
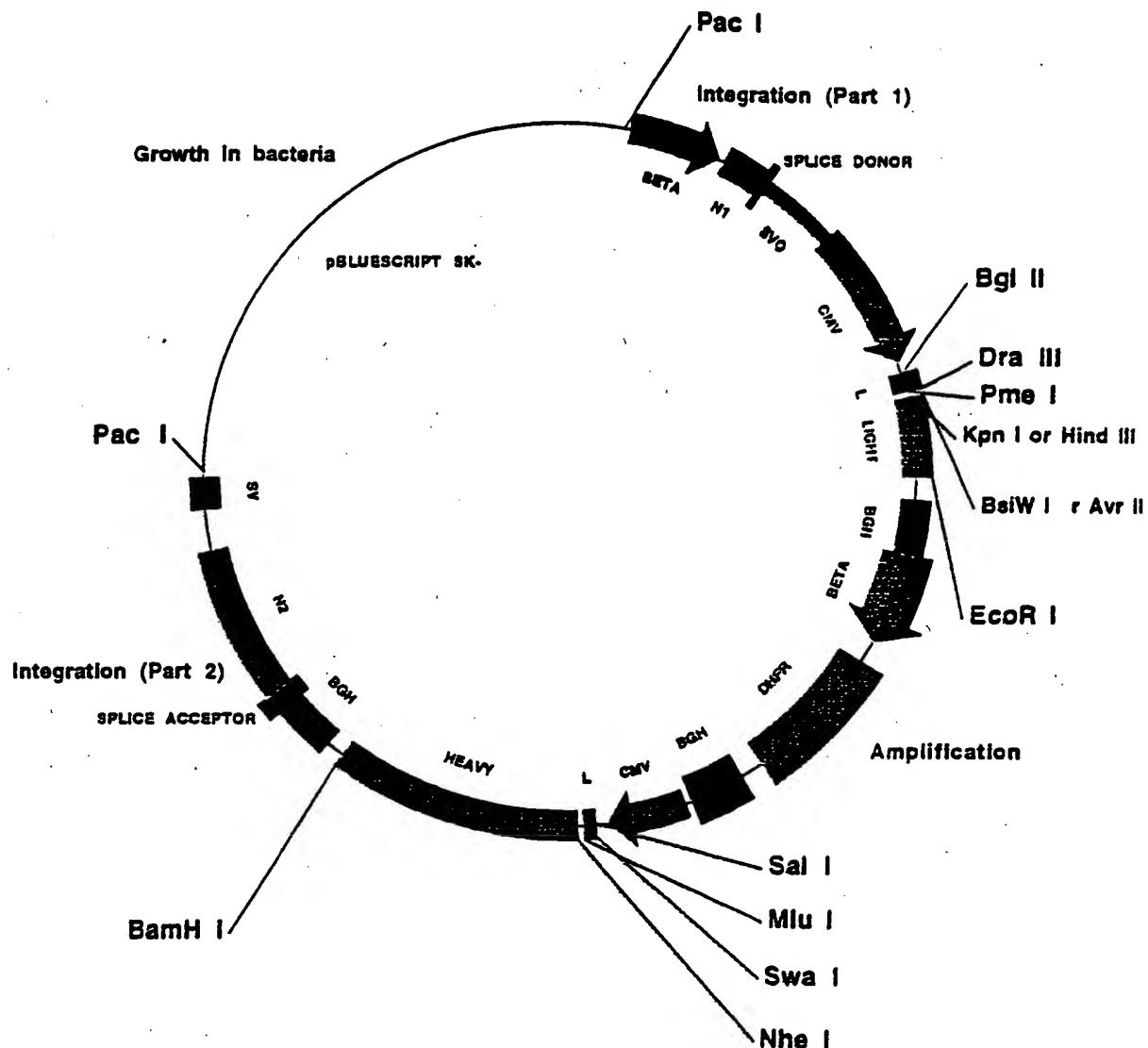


FIGURE 2

NEOSPLA



CMV = Cytomegalovirus promoter/enhancer
 BETA = Mouse Beta globin major promoter
 SVO = SV40 origin
 BGH = Bovine growth hormone polyadenylation
 SV = SV40-polyadenylation
 N1 = Neomycin phosphotransferase exon 1
 N2 = Neomycin phosphotransferase exon 2
 LIGHT = Human immunoglobulin kappa or lambda constant region
 DHFR = Dihydrofolate Reductase
 HEAVY = Human immunoglobulin gamma 1 or gamma 4 PE constant region
 L = Leader

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FIGURE 3a

Length of 7C10 Light/Primatized: 705 bp; Listed from: 1 to: 705;
 Translated from: 1 to: 703 (Entire region);
 Genetic Codes used: Universal; Fri, May 26, 1995 11:11 AM

Frame 1 M R V P A Q L L G L L L W L P G A R
 ATG AGG GTC CCC GCT CAG CTC CTG GGG CTC CTG CTG CTG TGG CTC CCA GGT GCA CGA
 9 18 27 36 45 54

C A Y E L T Q P P S V S V S P G Q T A R I
 TGT GCC TAT GAA CTG ACT CAG CCA CCC TCG GTG TCA GTG TCC CCA GGA CAG ACG GCC AGG ATC
 63 72 81 90 99 108 117

T C G G D N S R N E Y V H W Y Q Q K P A R
 ACC TGT GGG GCA GAC AAC AGT AGA AAT GAA TAT GTC CAC TGG TAC CAG CAG AAG CCA GCG CGG
 126 135 144 153 162 171 180

A P I L V I Y D D S D R P S G I P E R F S
 GCC CCT ATA CTG GTC ATC TAT GAT GAT AGT GAC CGG CCC TCA GGG ATC CCT GAG CGA TTC TCT
 189 198 207 216 225 234 243

G S K S G N T A T L T I N G V E A G D E A
 GGC TCC AAA TCA GGG AAC ACC GCC ACC CTG ACC ATC AAC GGG GTC GAG GCC GGG GAT GAG GCT
 252 261 270 279 288 297 306

D Y Y C Q V W D R A S D H P V F G G G T R
 GAC TAT TAC TGT CAG GTG TGG GAC AGG GCT GAT CAT CCG GTC TTC GGA GGA GGG ACC CGG
 315 324 333 342 351 360 369

V T V L G Q P K A A P S V T L F P P S S E
 GTG ACC GTC CTA GGT CAG CCC AAG GCT GCC CCC TCG GTC ACT CTG TTC CCG CCC TCC TCT GAG
 378 387 396 405 414 423 432

E L Q A N K A T L V C L I S D F Y P G A V
 GAG CTT CAA GCC AAC AAG GCC ACA CTG GTG TGT CTC ATA AGT GAC TTC TAC CCG GGA GCC GTG
 441 450 459 468 477 486 495

T V A W K A D S S P V K A G V E T T T P S
 ACA GTG GCC TGG AAG GCA GAT AGC AGC CCC GTC AAG GCG GGA GTG GAG ACC ACC ACA CCC TCC
 504 513 522 531 540 549 558

K Q S N N K Y A A S S Y L S L T P E Q W K
 AAA CAA AGC AAC AAC AAG TAC GCG GCC AGC AGC TAC CTG AGC CTG AGC CCT GAG CAG TGG AAG
 567 576 585 594 603 612 621

S H R S Y S C Q V T H E G S T V E K T V A
 TCC CAC AGA AGC TAC AGC TGC CAG GTC ACG CAT GAA GGG AGC ACC GTG GAG AAG ACA GTG GCC
 630 639 648 657 666 675 684

P T E C S
 CCT ACA GAA TGT TCA TGA
 693 702

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FIGURE 3b

Length of 7C10 heavy/Primateized: 1431 bp; Listed from: 1 to: 1431;
 Translated from: 1 to: 1429 (Entire region);
 Genetic Code used: Universal; Fri, May 26, 1995 11:11 AM

Frame 1 M K H L W F F L L L V A A P R W V L S
 ATG AAA CAC CTG TGG TTC CTC CTC CTG GTG GCA GCT CCC AGA TGG GTC CTG TCC
 9 18 27 36 45 54

Q V K L Q Q W G E G L L Q P S E T L S R T.
 CAG GTG AAG CTG CAG CAG TGG GGC GAA GGA CTT CTG CAG CCT TCG GAG ACC CTG TCC CGC ACC
 63 72 81 90 99 108 117

C V V S G G S I S G Y Y Y W T W I R Q T P
 TGC GTT GTC TCT GGT GGC TCC ATC AGC GGT TAC TAC TAC TGG ACC TGG ATC CGC CAG ACC CCA
 126 135 144 153 162 171 180

G R G L E W I G H I Y G N G A T T N Y N P
 GGG AGG GGA CTG GAG TGG ATT GGC CAT ATT TAT GGT AAT GGT GCG ACC ACC AAC TAC AAT CCC
 189 198 207 216 225 234 243

S L K S R V T I S K D T S K N Q F F L N L
 TCC CTC AAG AGT CGA GTC ACC ATT TCA AAA GAC ACG TCC AAG AAC CAG TTC TTC CTG AAC TTG
 252 261 270 279 288 297 306

N S V T D A D T A V Y Y C A R G P R P D C
 AAT TCT GTG ACC GAC GCG GAC ACG GCC GTC TAT TAC TGT GCG AGA GGC CCT CGC CCT GAT TGC
 315 324 333 342 351 360 369

T T I C Y G G W V D V W G P G D L V T V S
 ACA ACC ATT TGT TAT GGC GGC TGG GTC GAT GTC TGG GGC CCG GGA GAC CTG GTC ACC GTC TCC
 378 387 396 405 414 423 432

S A S T K G P S V F P L A P S S K S T S G
 TCA GCT AGC ACC AAG GGC CCA TCG GTC TTC CCC CTG GCA CCC TCC TCC AAG AGC ACC TCT GGG
 441 450 459 468 477 486 495

G T A A L G C L V K D Y F P E P V T V S W
 GGC ACA GCG GCC CTG GGC TGC CTG GTC AAG GAC TAC TTC CCC GAA CGG GTG ACG GTG TCG TGG
 504 513 522 531 540 549 558

N S G A L T S G V H T F P A V L Q S S G L
 AAC TCA GGC GCC CTG ACC AGC GGC GTG CAC ACC TTC CCG GCT GTC CTA CAG TCC TCA GGA CTC
 567 576 585 594 603 612 621

Y S L S S V V T V P S S S L G T Q T Y I C
 TAC TCC CTC AGC AGC GTG GTG ACC GTG CCC TCC AGC AGC TTG GGC ACC CAG ACC TAC ATC TGC
 630 639 648 657 666 675 684

N V N H K P S N T K V D K K A E P K S C D
 AAC GTG AAT CAC AAG CCC AGC AAC ACC AAG GTG GAC AAG AAA GCA GAG CCC AAA TCT TGT GAC
 693 702 711 720 729 738 747

K T H T C P P C P A P E L L G G P S V F L
 AAA ACT CAC ACA TGC CCA CCG TGC CCA GCA CCT GAA CTC CTG GGG GGA CGG TCA GTC TTC CTC
 756 765 774 783 792 801 810

P P K P K D T L M I S R T P E V T C V V
 TTC CCC CCA AAA CCC AAG GAC ACC CTC ATG ATC TCC CGG ACC CCT GAG GTC ACA TGC GTG GTG
 819 828 837 846 855 864 873

D V S H E D P E V K F N W Y V D G V E V
 GTG GAC GTG AGC CAC GAA GAC CCT GAG GTC AAG TTC AAC TGG TAC GTG GAC GGC GTG GAG GTG
 882 891 900 909

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FIGURE 3b (Continued)

H	N	A	K	T	K	P	R	E	E	Q	Y	N	S	T	Y	R	V	V	V	S	V
CAT	AAT	GCC	AAG	ACA	AAG	CCG	CGG	GAG	GAG	CAG	TAC	AAC	AGC	ACG	TAC	CCT	GTC	GTC	GTC	AGC	GTC
945	954					963				972			981			990				999	
L	T	V	L	H	Q	D	W	L	N	G	K	E	Y	K	C	K	V	S	N	K	
CTC	ACC	GTC	CTG	CAC	CAG	GAC	TGG	CTG	AAT	GCC	AAG	GAG	TAC	AAG	TGC	AAG	GTC	TCC	AAC	AAA	
1008	1017						1026			1035			1044			1053			1062		
A	L	P	A	P	I	E	K	T	I	S	K	A	K	G	Q	P	R	E	P	Q	
GCC	CTC	CCA	GCC	CCC	ATC	GAG	AAA	ACC	ATC	TCC	AAA	GCC	AAA	GCG	CAG	CCC	CGA	GAA	CCA	CAG	
1071	1080					1089				1098			1107			1116			1125		
V	Y	T	L	P	P	S	R	D	E	L	T	K	N	Q	V	S	L	T	C	L	
GTC	TAC	ACC	CTG	CCC	CCA	TCC	CGG	GAT	GAG	CTG	ACC	AAG	AAC	CAG	GTC	AGC	CTG	ACC	TGC	CTG	
1134	1143					1152				1161			1170			1179			1188		
V	K	G	F	Y	P	S	D	I	A	V	E	W	E	S	N	G	Q	P	E	N	
GTC	AAA	GGC	TTC	TAT	CCC	AGC	GAC	ATC	GCC	GTC	GAG	TGG	GAG	AGC	AAT	GGG	CAG	CCG	GAG	AAC	
1197	1206					1215				1224			1233			1242			1251		
N	Y	K	T	T	P	P	V	L	D	S	D	G	S	F	F	L	Y	S	K	L	
AAC	TAC	AAG	ACC	ACG	CCT	CCC	GTG	CTG	GAC	TCC	GAC	GCG	TCC	TTC	TTC	CTC	TAC	AGC	AAG	GTC	
1260	1269					1278			1287			1296			1305			1314			
T	V	D	K	S	R	W	Q	Q	G	N	V	F	S	C	S	V	M	H	E	A	
ACC	GTG	GAC	AAG	AGC	AGG	TGG	CAG	CAG	GGG	AAC	GTC	TTC	TCA	TGC	TCC	GTG	ATG	CAT	GAG	GCT	
1323	1332					1341				1350			1359			1368			1377		
L	H	N	H	Y	T	Q	K	S	L	S	L	S	P	G	K	.					
CTG	CAC	AAC	CAC	TAC	ACG	CAG	AAG	AGC	CTC	TCC	CTG	TCT	CCG	GGT	AAA	TGA					
1386	1395					1404				1413			1422			1431					

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FIGURE 4a

Length of 7B6 light/Primatized: 720 bp; Listed from: 1 to: 720;
 Translated from: 1 to: 718 (Entire region);
 Genetic Code used: Universal; Fri, May 26, 1995 11:10 AM

Frame 1 M S L P A Q L L G L L L L C V P G S S
 ATG AGC CTC CCT GCT CAG CTC CTC GGG CTG CTA TTG CTC TGC GTC CCC GGG TCC AGT
 9 18 27 36 45 54

G E V V M T Q S P L S L P I T P G E P A S
 GGG GAA GTT GTG ATG ACT CAG TCT CCA CTG TCC CTT CCT ATC ACA CCT GGA GAG CCG GCC TCC
 63 72 81 90 99 108 117

I S C R S S Q S L K H S N G D T F L S W Y
 ATC TCC TGT AGG TCT AGT CAA AGC CTT AAA CAC AGT AAT GGA GAC ACC TTC CTG AGT TGG TAT
 126 135 144 153 162 171 180

Q Q K P G Q P P R L L I Y K V S N R D S G
 CAG CAG AAG CCA GGC CAA CCT CCA AGG CTC CTG ATT TAT AAG GTT TCT AAC CCG GAC TCT GGG
 189 198 207 216 225 234 243

V P D R F S G S G A G T D F T L K I S A V
 GTC CCA GAC AGA TTC AGC GGC AGT GGG GCA GGG ACA GAT TTC ACA CTG AAA ATC AGC GCA GTG
 252 261 270 279 288 297 306

E A E D V G V Y F C G Q G T R T P P T F G
 GAG GCT GAA GAT GTT GGG GTT TAT TTC TGC GGG CAA GGT ACA AGG ACT CCT CCC ACT TTC GGC
 315 324 333 342 351 360 369

G G T K V E I K R T V A A P S V F I F P P
 GGA GGG ACC AAG GTG GAA ATC AAA CGT ACG GTG GCT GCA CCA TCT GTC TTC ATC TTC CCG CCA
 378 387 396 405 414 423 432

S D E Q L K S G T A S V V C L L N N F Y P
 TCT GAT GAG CAG TTG AAA TCT GGA ACT GCC TCT GTT GTG TGC CTG CTG AAT AAC TTC TAT CCC
 441 450 459 468 477 486 495

R E A K V Q W K V D N A L Q S G N S Q E S
 AGA GAG GCC AAA GTA CAG TGG AAG GTG GAT AAC GCC CTC CAA TCG GGT AAC TCC CAG GAG AGT
 504 513 522 531 540 549 558

V T E Q D S K D S T Y S L S S T L T L S K
 GTC ACA GAG CAG GAC AGC AAG GAC AGC ACC TAC AGC CTC AGC AGC ACC CTG AGC CTG AGC AAA
 567 576 585 594 603 612 621

A D Y E K H K V Y A C E V T H Q G L S S P
 GCA GAC TAC GAG AAA CAC AAA GTC TAC GCC TGC GAA GTC ACC CAT CAG GGC CTG AGC TCG CCC
 630 639 648 657 666 675 684

V T K S F N R G E C .
 GTC ACA AAG AGC TTC AAC AGG GGA GAG TGT TGA
 693 702 711 720

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FIGURE 4b

Length of 7B6 heavy/Primatized: 1437 bp; Listed from: 1 to: 1437;
 Translated from: 1 to: 1435 (Entire region);
 Genetic Code used: Universal; Fri, May 26, 1995 11:09 AM

gamma 1	M	G	W	S	L	I	L	L	F	L	V	A	V	A	T	R	V	Q	C		
	ATG	GGT	TGG	ACC	CTC	ATC	TTG	CTC	TTC	CCT	GTC	GCT	GTT	GCT	ACG	CGT	GTC	CAG	TGT		
	9	18	27	36	45	54															
E	V	Q	L	V	E	S	G	G	G	L	V	Q	P	G	G	S	L	R	V	S	
	GAG	GTC	CAG	CTG	GTC	GAG	TCT	GGG	GGG	GGC	TTG	GTC	CGG	CCT	GGC	GGG	TCC	CTG	AGA	GTC	TCC
	63	72	81	90	99	108														117	
C	A	V	S	G	F	T	F	S	D	H	Y	M	Y	W	F	R	Q	A	P	G	
	TGT	GCA	GTC	TCT	GGA	TTC	ACC	TTC	AGT	GAC	CAC	TAC	ATG	TAT	TGG	TTC	CGC	CGG	GCT	CCA	GGG
	126	135	144	153	162	171														180	
K	G	P	E	W	V	G	F	I	R	N	K	P	N	G	G	T	T	E	Y	A	
	AAG	GGG	CCG	GAA	TGG	GTA	GGT	TTC	ATT	AGA	AAC	AAA	CCG	AAC	GGT	GGG	ACA	ACA	GAA	TAC	GCC
	189	198	207	216	225	234														243	
A	S	V	K	D	R	F	T	I	S	R	D	D	S	K	S	I	A	Y	L	Q	
	CGG	TCT	GTG	AAA	GAC	AGA	TTC	ACC	ATC	TCC	AGA	GAT	GAT	TCC	AAA	AGC	ATC	GCC	TAT	CTG	CAA
	252	261	270	279	288	297														306	
M	S	S	L	K	I	E	D	T	A	V	Y	Y	C	T	T	S	Y	I	S	H	
	ATG	AGC	AGC	CTG	AAA	ATC	GAG	GAC	ACG	GCC	GTC	TAT	TAC	TGT	ACT	ACA	TCC	TAC	ATT	TCA	CAT
	315	324	333	342	351	360														369	
C	R	G	G	V	C	Y	G	G	Y	F	E	F	W	G	Q	G	A	L	V	T	
	TGT	CGG	GGT	GGT	GTC	TGC	TAT	GGA	GGT	TAC	TTC	GAA	TTC	TGG	GGC	CGG	GCC	CTG	GTC	ACC	
	378	387	396	405	414	423														432	
V	S	S	A	S	T	K	G	P	S	V	F	P	L	A	P	S	S	K	S	T	
	GTC	TCC	TCA	GCT	AGC	ACC	AAG	GGC	CCA	TCG	GTC	TTC	CCC	CTG	GCA	CCC	TCC	TCC	AAG	AGC	ACC
	441	450	459	468	477	486														495	
S	G	G	T	A	A	L	G	C	L	V	K	D	Y	F	P	E	P	V	T	V	
	TCT	GGG	GGC	ACA	GGC	GCC	CTG	GGC	TGC	CTG	GTC	AAG	GAC	TAC	TTC	CCC	GAA	CCG	GTG	ACG	GTG
	504	513	522	531	540	549														558	
S	W	N	S	G	A	L	T	S	G	V	H	T	F	P	A	V	L	Q	S	S	
	TCG	TGG	AAC	TCA	GGC	GCC	CTG	ACC	AGC	GGC	GTG	CAC	ACC	TTC	CCG	GCT	GTC	CTA	CAG	TCC	TCA
	567	576	585	594	603	612														621	
G	L	Y	S	L	S	S	V	V	T	V	P	S	S	S	L	G	T	Q	T	Y	
	GGA	CTC	TAC	TCC	CTC	AGC	AGC	GTG	GTG	ACC	GTG	CCC	TCC	AGC	AGC	TTG	GGC	ACC	CAG	ACC	TAC
	630	639	648	657	666	675														684	
I	C	N	V	N	H	K	P	S	N	T	K	V	D	K	K	A	E	P	K	S	
	ATC	TGC	AAC	GTG	AAT	CAC	AAG	CCC	AGC	AAC	ACC	AAG	GTG	GAC	AAG	AAA	GCA	GAG	CCC	AAA	TCT
	693	702	711	720	729	738														747	
C	D	K	T	H	T	C	P	P	C	P	A	P	E	L	L	G	G	P	S	V	
	TGT	GAC	AAA	ACT	CAC	ACA	TGC	CCA	CCG	TGC	CCA	GCA	CCT	GAA	CTC	CTG	GGG	GGG	CCG	TCA	GTC
	756	765	774	783	792	801														810	
F	L	F	P	P	K	P	K	D	T	L	M	I	S	R	T	P	E	V	T	C	
	TTC	CTC	TTC	CCC	CCA	AAA	CCC	AAG	GAC	ACC	CTC	ATG	ATC	TCC	CGG	ACC	CCT	GAG	GTC	ACA	TGC
	819	828	837	846	855	864														873	
V	V	V	D	V	S	H	E	D	P	E	V	K	F	N	W	Y	V	D	G	V	
	GTG	GTG	GAC	GTG	AGC	CAC	GAA	GAC	CCT	GAG	GTC	AAG	TTC	AAC	TGG	TAC	GTG	GAC	GGC	GTG	
	882	891	900	909	918	927														936	

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FIGURE 4b (Continued)

E	V	H	N	A	K	T	K	P	R	E	E	Q	Y	N	S	T	Y	R	V	V
GAG	GTG	CAT	AAT	GCC	AAG	ACA	AAG	CCG	GGG	GAG	GAG	CAG	TAC	AAC	AGC	ACG	TAC	CGT	GTG	GTC
945	954			963				972				981			990			999		
S	V	L	T	V	L	H	Q	D	W	L	N	G	K	E	Y	K	C	K	V	S
AGC	GTC	CTC	ACC	GTC	CTG	CAC	CAG	GAC	TGG	CTG	AAT	GCC	AAG	GAG	TAC	AAG	TGC	AAG	GTC	TCC
1008	1017			1026				1035				1044			1053			1062		
N	K	A	L	P	A	P	I	E	R	T	I	S	K	A	K	G	Q	P	R	E
AAC	AAA	GCC	CTC	CCA	GCC	CCC	ATC	GAG	AAA	ACC	ATC	TCC	AAA	GCC	AAA	GGG	CAG	CCC	CGA	GAA
1071	1080			1089				1098				1107			1116			1125		
P	Q	V	Y	T	L	P	P	S	R	D	E	L	T	K	N	Q	V	S	L	T
CCA	CAG	GTG	TAC	ACC	CTG	CCC	CCA	TCC	CGG	GAT	GAG	CTG	ACC	AAG	AAC	CAG	GTC	AGC	CTG	ACC
1134	1143			1152				1161				1170			1179			1188		
C	L	V	K	G	F	Y	P	S	D	I	A	V	E	W	E	S	N	G	Q	P
TGC	CTG	GTC	AAA	GCC	TTC	TAT	CCC	AGC	GAC	ATC	GCC	GTG	GAG	TGG	GAG	AGC	AAT	GGG	CAG	CCG
1197	1206			1215				1224				1233			1242			1251		
E	N	N	Y	K	T	T	P	P	V	L	D	S	D	G	S	F	F	L	Y	S
GAG	AAC	AAC	TAC	AAG	ACC	ACG	CCT	CCC	GTG	CTG	GAC	TCC	GAC	GCC	TCC	TTC	TTC	CTC	TAC	AGC
1260	1269			1278				1287				1296			1305			1314		
K	L	T	V	D	K	S	R	W	Q	Q	G	N	V	F	S	C	S	V	M	H
AAG	CTC	ACC	GTG	GAC	AAG	AGC	AGG	TGG	CAG	CAG	GGG	AAC	GTC	TTC	TCA	TGC	TCC	GTG	ATG	CAT
1323	1332			1341				1350				1359			1368			1377		
E	A	L	H	N	H	Y	T	Q	K	S	L	S	L	S	P	G	K			
GAG	GCT	CTG	CAC	AAC	CAC	TAC	ACG	CAG	AAG	AGC	CTC	TCC	CTG	TCT	CCG	GGT	AAA	TGA		
1386	1395			1404				1413				1422			1431					

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FIGURE 5a

Length of 16C10 Lambda/Primatized : 711 bp;

Listed from: 1 to: 711;

Translated from: 1 to: 709 (Entire region);

Genetic Code used: Universal; Fri, May 26, 1995 11:08 AM

frame 1	M	R	V	P	A	Q	L	L	G	L	L	L	L	L	W	L	P	G	A	R
	ATG	AGG	GTC	CCC	GCT	CAG	CTC	CTG	GGG	CTC	CTG	CTG	CTG	TGG	CTC	CCA	GGT	GCA	CGA	
	9						18				27				36		45			54
C	E	S	V	L	T	Q	P	P	S	V	S	G	A	P	G	Q	K	V	T	I
IGT	GAG	TCT	GTC	CTG	ACA	CAG	CCG	CCC	TCA	GTG	TCT	GGG	GCC	CCA	GGG	CAG	AAG	GTC	ACC	ATC
63									81		90		99			108				117
S	C	T	G	S	T	S	N	I	G	G	Y	D	L	H	W	Y	Q	Q	L	P
TCG	TGC	ACT	GGG	AGC	ACC	TCC	AAC	ATT	GGA	GGT	TAT	GAT	CTA	CAT	TGG	TAC	CAG	CAG	CTC	CCA
126								135		144		153			162		171			180
G	T	A	P	K	L	L	I	Y	D	I	N	K	R	P	S	G	I	S	D	R
GGA	ACG	GCC	CCC	AAA	CTC	CTC	ATC	TAT	GAC	ATT	AAC	AAG	CGA	CCC	TCA	GGG	ATT	TCT	GAC	CGA
189								198		207		216			225		234			243
F	S	G	S	K	S	G	T	A	A	S	L	A	I	T	G	L	Q	T	E	D
TTC	TCT	GGC	TCC	AAG	TCT	GGT	ACC	GCG	GCC	TCC	CTG	GCC	ATC	ACT	GGG	CTC	CAG	ACT	GAG	GAT
252								261		270		279			288		297			306
E	A	D	Y	Y	C	Q	S	Y	D	S	S	L	N	A	Q	V	F	G	G	G
GAG	GCT	GAT	TAT	TAC	TGC	CAG	TCC	TAT	GAC	AGC	AGC	CTG	AAT	GCT	CAG	GTA	TTC	GGG	GGG	
315								324		333		342			351		360			369
T	R	L	T	V	L	G	Q	P	K	A	A	P	S	V	T	L	F	P	P	S
ACC	CGG	CTG	ACC	GTC	CTA	GGT	CAG	CCC	AAG	GCT	GCC	CCC	TCG	GTC	ACT	CTG	TTC	CCG	CCC	TCC
378								387		396		405			414		423			432
S	E	E	L	Q	A	N	K	A	T	L	V	C	L	I	S	D	F	Y	P	G
TCT	GAG	GAG	CTT	CAA	GCC	AAC	AAG	GCC	ACA	CTG	GTG	TGT	CTC	ATA	AGT	GAC	TTC	TAC	CCG	GGG
441								450		459		468			477		486			495
A	V	T	V	A	W	K	A	D	S	S	P	V	K	A	G	V	E	T	T	T
GCC	GTG	ACA	GTG	GCC	TGG	AAG	GCA	GAT	AGC	AGC	CCC	GTC	AAG	GCG	GGG	GTG	GAG	ACC	ACC	ACA
504								513		522		531			540		549			558
P	S	K	Q	S	N	N	K	Y	A	A	S	S	Y	L	S	L	T	P	E	Q
CCC	TCC	AAA	CAA	AGC	AAC	AAC	AAG	TAC	GCG	GCC	AGC	AGC	TAC	CTG	AGC	CTG	ACG	CCT	GAG	CAG
567								576		585		594			603		612			621
W	K	S	H	R	S	Y	S	C	Q	V	T	H	E	G	S	T	V	E	K	T
TCG	AAG	TCC	CAC	AGA	AGC	TAC	AGC	TGC	CAG	GTC	ACG	CAT	GAA	GGG	AGC	ACC	GTG	GAG	AAG	ACA
630								639		648		657			666		675			684
V	A	P	T	E	C	S														
GTG	GCC	CCT	ACA	GAA	TGT	TCA	TGA													
693								702		711										

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FIGURE 5b

Length of 16C10 heavy/primatized: 1431 bp;
 Listed from: 1 to: 1431;
 Translated from: 1 to: 1429 (Entire region);
 Genetic Code used: Universal; Fri, May 26, 1995 11:08 AM

frame 1	M	K	H	L	W	F	F	L	L	L	V	A	A	P	R	W	V	L	S		
	ATG	AAA	CAC	CTG	TGG	TTC	TTC	CTC	CTC	CTG	GTG	GCA	GCT	CCC	AGA	TGG	GTC	CTG	TCC		
	9	18	27	36	45														54		
2	V	Q	L	Q	E	S	G	P	G	L	V	K	P	S	E	T	L	S	L	T	
	CAG	GTG	CAG	CTG	CAG	GAG	TCG	GGC	CCA	GGA	CTG	GTG	AAG	CCT	TCG	GAG	ACC	CTG	TCC	CTC	ACC
	63	72	81	90	99															108	117
3	A	V	S	G	G	S	I	S	G	G	Y	G	W	G	W	I	R	Q	P	P	
	GGC	GCT	GTC	TCT	GGT	GGC	TCC	ATC	AGC	GGT	TAT	GGC	TGG	GGC	TGG	ATC	CAG	CCC	CCA		
	126	135	144	153	162														171	180	
4	G	K	G	L	E	W	I	G	S	F	Y	S	S	S	G	N	T	Y	Y	N	P
	GGG	AAG	GGG	CTG	GAG	TGG	ATT	GGG	AGT	TTC	TAT	AGT	AGT	AGT	GGG	AAC	ACC	TAC	TAC	AAC	CCC
	189	198	207	216	225														234	243	
5	S	L	K	S	Q	V	T	I	S	T	D	T	S	K	N	Q	F	S	L	K	L
	TCC	CTC	AAG	AGT	CAA	GTC	ACC	ATT	TCA	ACA	GAC	ACG	TCC	AAG	AAC	CAG	TTC	TCC	CTG	AAG	CTG
	252	261	270	279	288														297	306	
6	N	S	M	T	A	A	D	T	A	V	Y	Y	C	V	R	D	R	L	F	S	V
	AAC	TCT	ATG	ACC	GCC	GGG	GAC	ACG	GCC	GTG	TAT	TAC	TGT	GTG	AGA	GAT	CGT	CTT	TTT	TCA	GTG
	315	324	333	342	351														360	369	
7	V	G	M	V	Y	N	N	W	E	D	V	W	G	P	G	V	L	V	T	V	S
	GTG	GGA	ATG	GTG	TAC	AAC	AAC	TGG	TTC	GAT	GTC	TGG	GGC	CGG	GGG	GTC	CTG	GTC	ACC	GTC	TCC
	378	387	396	405	414														423	432	
8	S	A	S	T	K	G	P	S	V	F	P	L	A	P	S	S	K	S	T	S	G
	TCA	GCT	AGC	ACC	AAG	GGC	CCA	TCG	GTC	TTC	CCC	CTG	GCA	CCC	TCC	TCC	AAG	AGC	ACC	TCT	GGG
	441	450	459	468	477														486	495	
9	G	T	A	A	L	G	C	L	V	K	D	Y	F	P	E	P	V	T	V	S	W
	GGC	ACA	GCG	GCC	CTG	GGC	TGC	CTG	GTC	AAG	GAC	TAC	TTC	CCC	GAA	CGG	GTG	ACG	GTG	TGG	TGG
	504	513	522	531	540														549	558	
10	N	S	G	A	L	T	S	G	V	H	T	F	P	A	V	L	Q	S	S	G	L
	AAC	TCA	GCC	GCC	CTG	ACC	AGC	GGC	GTG	CAC	ACC	TTC	CGG	GCT	GTC	CTA	CAG	TCC	TCA	GGA	CTC
	567	576	585	594	603														612	621	
11	Y	S	L	S	S	V	V	T	V	P	S	S	S	L	G	T	Q	T	Y	I	C
	TAC	TCC	CTC	AGC	AGC	GTG	GTG	ACC	GTG	CCC	TCC	AGC	AGC	TTC	GGC	ACC	CAG	ACC	TAC	ATC	TGC
	630	639	648	657	666														675	684	
12	N	V	N	H	K	P	S	N	T	K	V	D	K	K	A	E	P	K	S	C	D
	AAC	GTG	AAT	CAC	AAG	CCC	AGC	AAC	ACC	AAG	GTG	GAC	AAG	AAA	GCA	GAG	CCC	AAA	TCT	TGT	GAC
	693	702	711	720	729														738	747	
13	K	T	H	T	C	P	P	C	P	A	P	E	L	L	G	G	P	S	V	F	L
	AAA	ACT	CAC	ACA	TGC	CCA	CCG	TGC	CCA	GCA	CCT	GAA	CTC	CTG	GGG	GGG	TCA	GTC	TTC	TTC	CTC
	756	765	774	783	792														801	810	
14	F	P	P	K	P	K	D	T	L	M	I	S	R	T	P	E	V	T	C	V	V
	TTC	CCC	CCA	AAA	CCC	AAG	GAC	ACC	CTC	ATG	ATC	TCC	CGG	ACC	CCT	GAG	GTC	ACA	TGC	GTG	GTG
	819	828	837	846	855														864	873	
15	V	D	V	S	H	E	D	P	E	V	K	F	N	W	Y	V	D	G	V	E	V
	GTG	GAC	GTG	AGC	CAC	GAA	GAC	CCT	GAG	GTC	AAG	TTC	AAC	TGG	TAC	GTG	GAC	GCC	GTG	GAG	GTG

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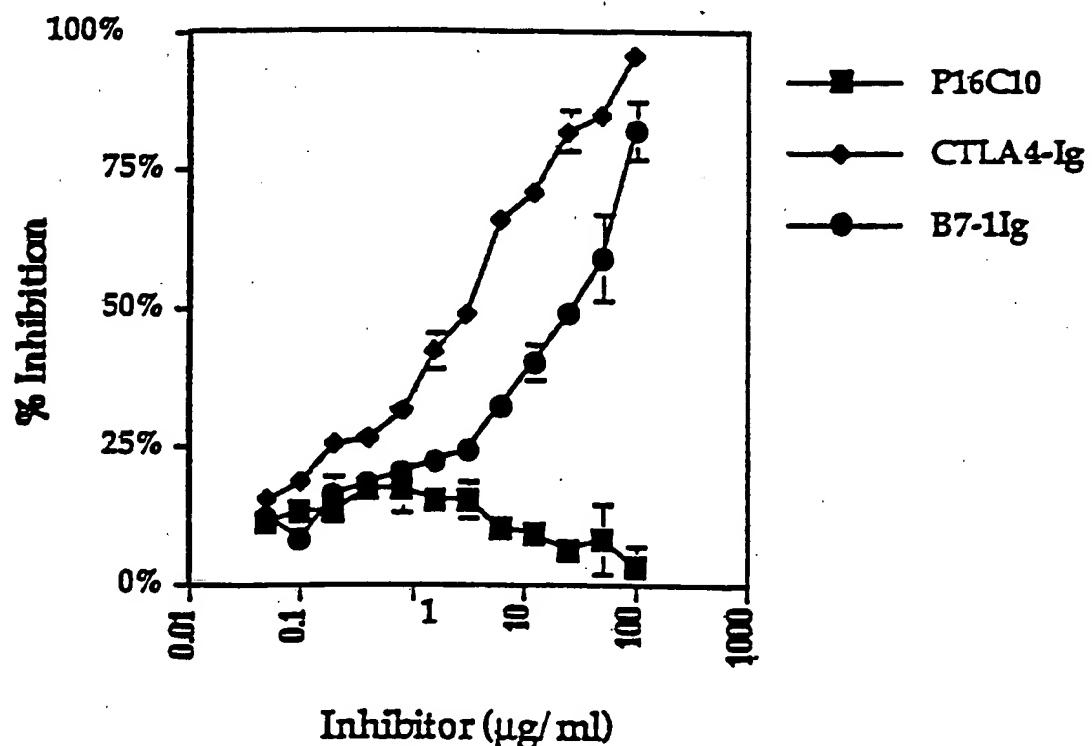
FIGURE 5b (Continued)

882	891	900	909	918	927	936
I N A K T K P R E E Q Y N S T Y R V V S V						
CAT AAT GCC AAG ACA AAG CCG CGG GAG GAG CAG TAC AAC AGC ACG TAC CGT GTG GTC AGC GTC						
945 954 963 972 981 990 999						
T V L H Q D W L N G K E Y K C K V S N K						
CTC ACC GTC CTG CAC CAG GAC TCG CTG AAT GGC AAG GAG TAC AAC TGC AAG GTC TCC AAC AAA						
1008 1017 1026 1035 1044 1053 1062						
A L P A P I E K T I S K A K G Q P R E P Q						
GCC CTC CCA GCC CCC ATC GAG AAA ACC ATC TCC AAA GCC AAA GGG CAG CCC CGA GAA CCA CAG						
1071 1080 1089 1098 1107 1116 1125						
V Y T L P P S R D E L T K N Q V S L T C L						
GTC TAC ACC CTG CCC CCA TCC CGG GAT GAG CTG ACC AAC CAG GTC AGC CTG ACC TGC CTG						
1134 1143 1152 1161 1170 1179 1188						
V K G F Y P S D I A V E W E S N G Q P E N						
GTC AAA GGC TTC TAT CCC AGC GAC ATC GCC GTG GAG TGG GAG AGC AAT GGG CAG CCG GAG AAC						
1197 1206 1215 1224 1233 1242 1251						
N Y K T T P P V L D S D G S F F L Y S K L						
AAC TAC AAG ACC ACG CCT CCC GTG CTG GAC TCC GAC GGC TCC TTC TTC CTC TAC AGC AAG CTC						
1260 1269 1278 1287 1296 1305 1314						
T V D K S R W Q Q G N V F S C S V M H E A						
ACC GTG GAC AAG AGC AGG TGG CAG CAG GGG AAC GTC TTC TCA TGC TCC GTG ATG CAT GAG GCT						
1323 1332 1341 1350 1359 1368 1377						
L H N H Y T Q K S L S L S P G K						
CTG CAC AAC CAC TAC AGC CAG AAG AGC CTC TCC CTG TCT CCG GGT AAA TGA						
1386 1395 1404 1413 1422 1431						

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Figure 6

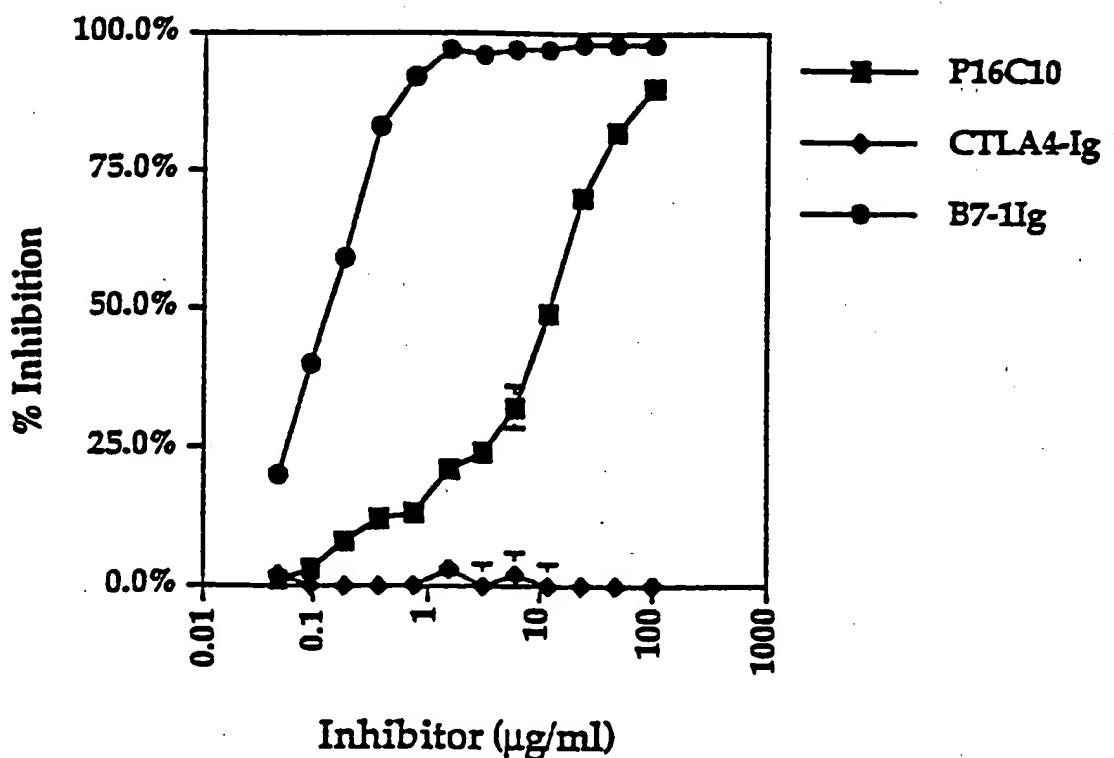
Competitive Binding of CTLA-4Ig-Biotin to B7-1 Transfected CHO Cells
in Presence of P16Cl0 MAb, CTLA-4Ig and B7-1Ig Fusion Proteins



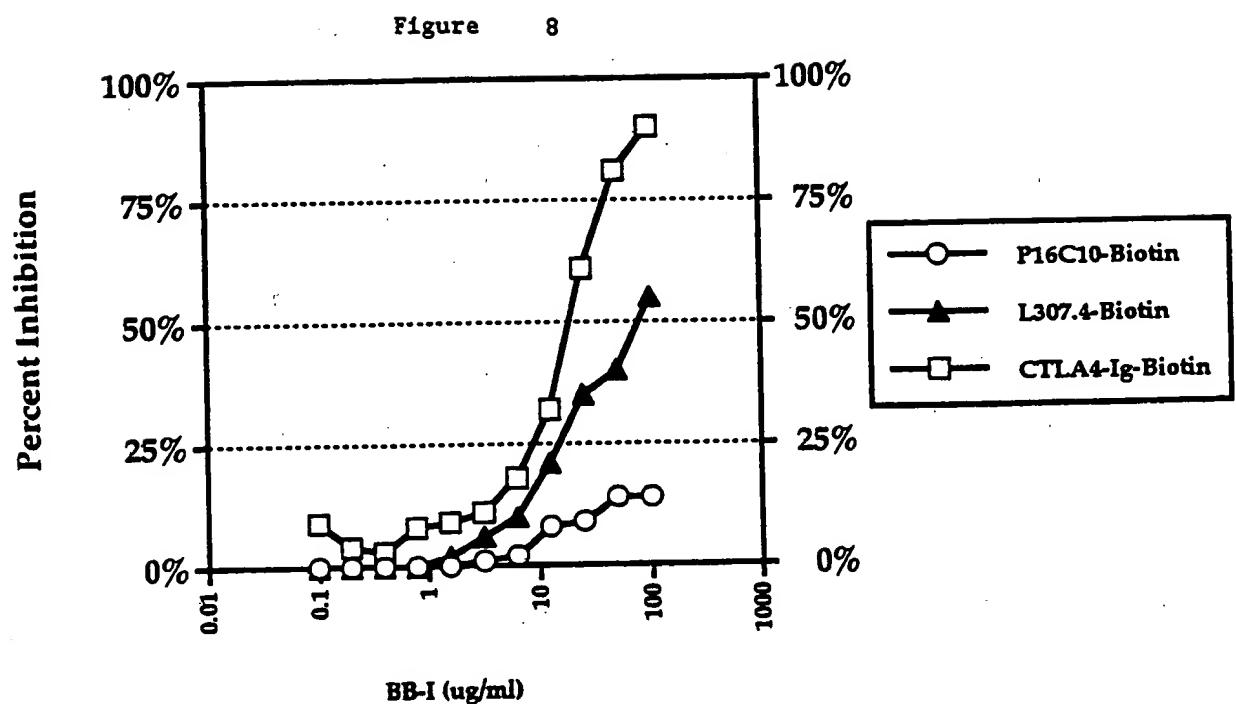
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Figure 7

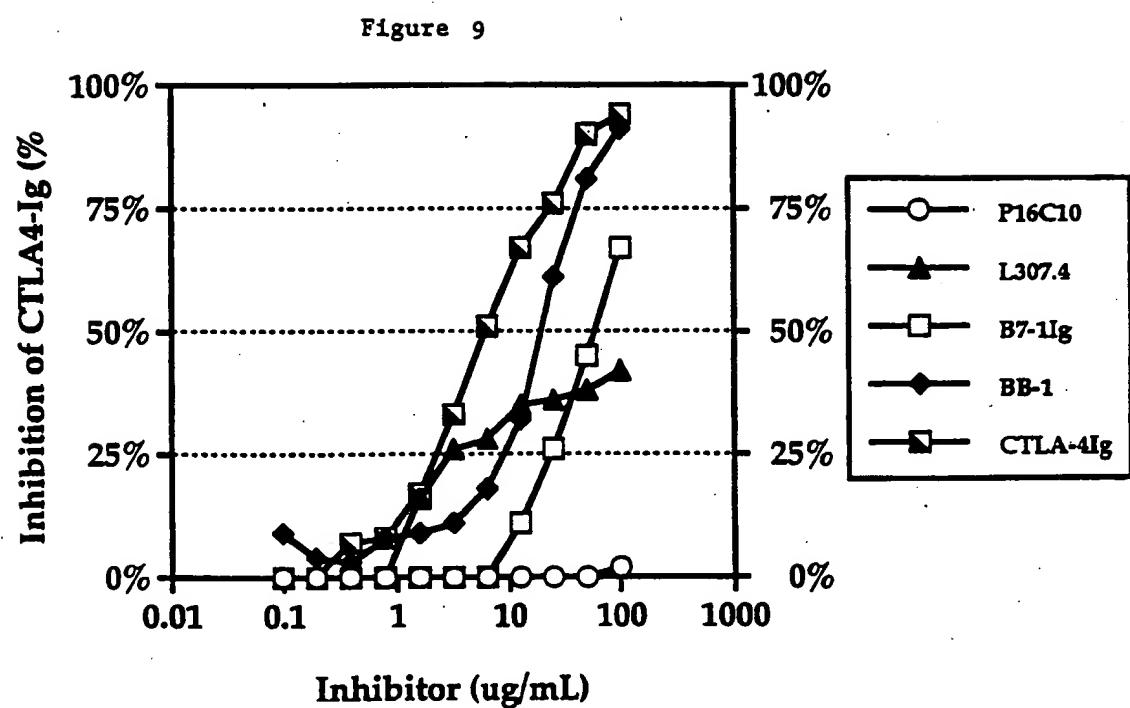
**Competitive Binding of P16C10-Biotin to B7-1 Transfected CHO Cells
in Presence of P16C10 MAb, CTLA-4Ig and B7-1Ig Fusion Proteins**



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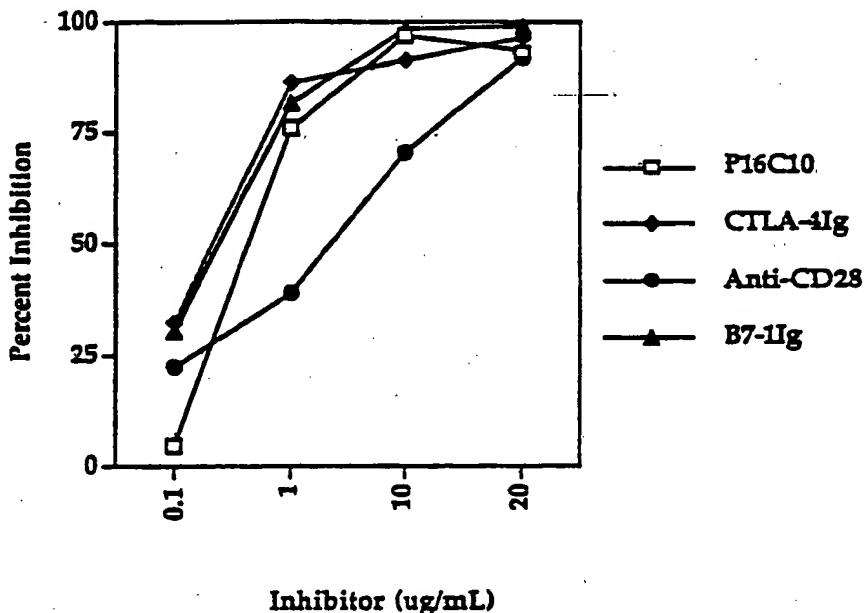
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Figure 10

Competitive Inhibition of Radiolabeled B7-1Ig Binding to CD28 Receptors on Isolated Non-Activated Peripheral Blood T Cells using Unlabeled Binding Proteins Recognizing Either B7-1 or CD28



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/19906

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : A61K 39/395; C07K 16/18, 16/28

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/1301., 142.1, 143.1, 144.1, 153.1, 154.1, 173.1; 530/387.1, 387.3, 388.1, 388.15, 388.22, 388.7, 388.73, 388.75

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

NONE

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG, BIOSIS, CA, EMBASE, MEDLINE, WPI
search terms: b7.1, ciba4, cd28, graft?, transplant?, gvhd

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	LINSLEY et al. Binding of the B Cell Activation Antigen B7 to CD28 Costimulates T Cell Proliferation and Interleukin 2 mRNA Accumulation. J. Exp. Med. March 1991, Volume 173, pages 721-730, see entire document.	1-3, 6, 7, 9-14, 16, 18, 26
Y	LINSLEY et al. T-Cell Antigen CD28 Mediates Adhesion With B Cells by Interacting with Activation Antigen B7/BB-1. Proc. Natl. Acad. Sci. USA. July 1990, Volume 87, pages 5031-5035, see entire document.	1-3, 6, 7, 9-14, 16, 18, 26
Y	US 5,434,131 A (P.S. LINSLEY et al.) 18 July 1995, see entire document.	1-3, 6, 7, 9-14, 16, 18, 26

 Further documents are listed in the continuation of Box C.

See patent family annex.

• Special categories of cited documents:		
•A• document defining the general state of the art which is not considered to be of particular relevance	•T•	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
•B• earlier document published on or after the international filing date	•X•	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
•L• document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	•Y•	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
•O• document referring to an oral disclosure, use, exhibition or other means	•Z•	document member of the same patent family
•P• document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

07 JANUARY 1998

Date of mailing of the international search report

03 FEB 1998

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231
Facsimile No. (703) 305-3230

Authorized officer

PHILLIP GAMBEL

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/19906

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,521,288 A (P.S. LINSLEY et al.) 28 May 1996, see entire document.	1-3, 6, 7, 9-14, 16, 18, 26

Form PCT/ISA/210 (continuation of second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/19906

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-3, 6, 7, 9-14, 16, 18, 26

Remark on Protest

The additional search fees were accompanied by the applicant's protest.
 No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)★

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/19906

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

424/1301., 142.1, 143.1, 144.1, 153.1, 154.1, 173.1; 530/387.1, 387.3, 388.1, 388.15, 388.22, 388.7, 388.73, 388.75

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This International Search Authority has found 28 inventions claimed in the International Application covered by the claims indicated below and it considers that the International Application does not comply with the requirements of Unity of Invention (Rules 13.1, 13.2 and 3.3 for the reasons indicated below:

This application contains claims directed to the following distinct species of the claimed invention; wherein the antibody specificity is:

- A) B7.1, claims 1-3, 6, 7, 9-14, 16, 18, 20, 22, 24, 26, 27 or
- B) B7.2., claims 1, 4-6, 8-10, 12, 13, 15, 17, 19, 21, 23, 25, 26, 28

These species do not share a common technical feature as these molecules differ in their structures and modes of action; therefore they are distinct.

In addition, this application contains claims directed to the following distinct species of the claimed invention and the specificity of B7.1 or B7.2; wherein the disease is:

- A) thrombocytopenia, claims 20, 21
- B) lupus, claims 20, 21
- C) diabetes, claims 20, 21
- D) arthritis, claims 20, 21
- E) psoriasis, claims 20, 21
- F) anemia, claims 20, 21
- G) IBD, claims 20, 21
- H) allergy, claims 20, 21
- I) multiple sclerosis,
- J) GVHD, claims 22, 23
- K) B cell lymphoma, claims 24, 25
- L) infectious diseases, claims 24, 25
- M) inflammatory diseases, claims 24, 25 or
- N) autoimmunity, claims 18,19.

These species do not share a common technical feature as these pathological conditions differ in etiologies and therapeutic endpoints; therefore they are distinct.

Accordingly, the claims are not so linked by a special technical feature within the meaning of PCT Rule 13.2 so as to form a single inventive concept.



131/180 - (C) FILE CANCERLIT

STN GEN Caesar accession number : 1965

AN - 96606486 CANCERLIT

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DN - 96606486

TI - Inhibition of costimulatory molecules as a means of treating a murin
T-cell dependent B-cell lymphoma

AU - Wrone-Smith T

CS - Wayne State Univ.

SO - Diss Abstr Int [B], (1995). Vol. 56, No. 2, pp. 736.
ISSN: 0419-4217.

DT - (THESIS)

FS - ICDB

LA - English

EM - 199605

AB - This dissertation focuses on the role of costimulation in the CD4+ T
dependent SJL B-cell lymphoma model. It has been determined by
flow microfluorometry that the SJL lymphoma, RCS5, expresses
B7 costimulatory molecules, B7 -1 and B7 -2, and
expresses LFA-1 and its ligand, ICAM-1. Blocking B7 /CD28 or
LFA-1/ICAM-1 costimulatory pathways with anti- B7 or anti-LFA-1
monoclonal antibodies significantly decreases naive or
tumor-activated CD4+ T-cell proliferation to the tumor stimulus in v
Furthermore, blocking B7 - or LFA-1-mediated costimulation
significantly inhibits RCS5 tumor growth in vivo. (Full text availab
from University Microfilms International, Ann Arbor, MI, as Order No
AADAA-I9519980)

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